Plants have been used for the treatment of diseases all over the world before the advent of modern clinical drugs. Natural photochemical are known to contain substances that can be used for therapeutic purposes or as precursor for the synthesis of novel drugs. Nearly 50% modern drugs are of natural products origin and as such these natural products play an important role in drug development in pharmaceutical industry. Plants remain the most common source of antimicrobial agents (Bibitha et al., 2002 and Maghrani et al., 2005). Many aromatic plants have been used traditionally in folk medicine as well as to extend the shelf life of foods, showing inhibition against bacteria, fungi and yeast (Hulin et al., 1998). Biologically active compounds from natural sources have always been a great interest for scientists working in infectious diseases (Perumal Samy and Ignacimuthu, 2000). There is an essential need to discover new antimicrobial compounds with diverse chemical structures and novel mechanisms of action. Therefore, search for medicinal plants with potential secondary metabolites have been extensively investigated as a source of medicinal agents. Drug resistance is a serious global problem, and spread of resistance poses additional challenges for clinicians and the pharmaceutical industry.

Use of herbal medicines in the developed world continue to rise because they are rich source of novel drugs and their bioactive principles form the basis in medicine, nutraceuticals, pharmaceutical intermediates and lead compounds in synthetic drugs (De et al., 2002 and Ncube et al., 2008). Screening medicinal plants for biologically active compounds offers clues to develop newer antimicrobial agents. These compounds after possible chemical manipulation provide new and improved drugs to treat the infectious diseases (Natarajan et al., 2003 and Shah et al., 2006). Plant based products extracts are cheaper alternatives to the development of synthetic drugs. The plant-derived medicines are based upon the premise that they contain natural substances that can promote health and alleviate illness. So returns to natural substances are an absolute need of our time (Swayamjot et al., 2005; Kumar et al., 2007). In the last few years a number of studies have been conducted to verify the effectiveness of plant extracts against bacterial infections (Prashanth et al., 2006; Ung et al., 2010). Azadirachta Indica belongs to the family Meliaceae, commonly known as neem. It is used in traditional medicine as a source of many therapeutic agents. A. indica (leaf, bark and seed) are known to contain antibacterial, antifungal activities against different pathogenic microorganisms and antiviral activity against vaccinia, chikungunya, measles and coxsackie B viruses (Biswas et al., 2002).
Different parts of neem (leaf, bark and seed oil) have been shown to exhibit wide pharmacological activities including; antioxidant, antimalarial, antimutagenic, anticarcinogenic, antiinflammatory, antihyperglycaemic, antulcer and antidiabetic properties (Talwar et al., 1997). The biological activities are attributed to the presence of many bioactive compounds in different parts. The present study evaluated the individual and in combination growth inhibitory effect of 4 medicinal plant extracts against 4 bacteria.

**MATERIALS AND METHODS**

Plants were collected between the month of June and July 2015 in the Chidambaram area Tamil Nadu, India. Plant leaves were initially dried in an airconditioned, dehumidified room, then further dried in an oven at ca. 40°C for a total of seven days, and then finally ground to a fine powder. Antimicrobial activity test was determined by the Kirby-Bauer disc diffusion method (Bauer et al., 1996). The Antimicrobial activity was tested against isolated 4 bacterial strains. The medicinal plants of *Hibiscus rosasinensis, Azadirachta indica, Ficus religiosa and Ocimum sanctum* leaves extract were tested by the disc diffusion method. The extracts were prepared by reconstituting with aquous.

The test microorganisms were seeded into respective Mueller-Hinton agar medium by spread plate method 10 μl (10 cells/ml) with the 24h cultures of bacteria growth in Mueller-Hinton agar broth. After solidification the filter paper discs (5 mm in diameter) impregnated with the extracts were placed on test-organism-seeded plates, *Escherichia coli, Salmonella typhi, Staphylococcus aureus* and *Enterobacter aerogen* were used for antibacterial test. Erythromycin (10 μg mlG1) used as positive control. The antibacterial assay plates were incubated at 37°C for 24h. After incubation, the results were observed and measured the diameter of inhibition zone (mm) around the each well.

**RESULTS AND DISCUSSION**

The antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Escherichia coli* 14 mm (Mean value in Dia.), when compared with other medicinal plant extracts. *Ficus religiosa* plant extract was zone of inhibition 12 mm (Mean value in Dia.) against *Escherichia coli* and 11 mm (Mean value in Dia.) against *Salmonella typhii*, *Staphylococcus aureus* and *Enterobacter aerogen*. The *Enterobacter aerogen* was highly inhibition 8 mm (Mean value in Dia.) against *Ocimum sanctum* plant extract.

<table>
<thead>
<tr>
<th>Name of the Species</th>
<th>Zone of inhibition ( dia in mm)</th>
<th>S1. Hibiscus rosasinensis</th>
<th>S2. Azadirachta indica</th>
<th>S3. Ficus religiosa</th>
<th>S4. Ocimum sanctum</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Salmonella typhi</em></td>
<td></td>
<td>10.33 ± 0.58</td>
<td>10.33 ± 1.15</td>
<td>10.33 ± 2.33</td>
<td>9.67 ± 2.08</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td></td>
<td>13.00 ± 3.00</td>
<td>10.67 ± 1.53</td>
<td>10.00 ± 3.60</td>
<td>10.00 ± 3.60</td>
</tr>
<tr>
<td><em>E coli</em></td>
<td></td>
<td>11.67 ± 2.52</td>
<td>14.33 ± 2.52</td>
<td>12.00 ± 2.00</td>
<td>11.67 ± 2.52</td>
</tr>
<tr>
<td><em>Enterobacter aerogen</em></td>
<td></td>
<td>9.33 ± 0.58</td>
<td>10.67 ± 0.58</td>
<td>10.00 ± 4.58</td>
<td>8.67 ± 3.51</td>
</tr>
</tbody>
</table>

Plate 1. Antimicrobial activity test against isolated bacteria from infected Fresh water carp *Catla catla*
At the same time *Hibiscus rosasinensis* plant extract was highly sensitive 13 mm (Mean value in Dia.) against *Staphylococcus aureus*. The *Azadirachta indica* and *Ficus religiosa* medicinal plant extracts are having best control of antibacterial activity. Antimicrobial activity test against five different medicinal plants (leaves) extracts such as *Hibiscus rosasinensis, Azadirachta indica, Ficus religiosa and Ocimum sanctum* (Leaves) were tested against some pathogenic bacteria such as *Escherichia coli, Salmonella typhi, Staphylococcus aureus* and *Enterobacter aerogen*. The antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Escherichia coli*, when compared with other medicinal plant extracts. According to Abalaka et al. (2012) were studied the antibacterial effects of *A. indica* on the test organisms revealed that *P. aeruginosa* showed the highest zones of inhibition (mm) followed by *S. aureus* while *E. coli* had the least zone of inhibition (mm) at various extract concentrations of 500mg/ml, 50mg/ml and 5mg/ml. The extracts of *A. indica* showed a higher value of zones of inhibition on the tested organisms. In a similar study hexane and aqueous extract of *Azadirachta indica*, inhibited *Es-cherichia coli*, *P. aeruginosa*, *S. pyogenes* and *S.aureus* (El-Mahmood et al., 2010).

![Fig. 1. Antimicrobial activity test against isolated bacteria from infected carp *Catla catla*](image)

Methanol extract of the leaves of *Azadirachta indica* exhibited pronounced activity (28mm) against *Bacillus subtilis*, high activity (18mm) against the Gram positive *Staphylococcus aureus* and the Gram-negative organisms *Proteus vulgaris* (18 mm) and *Salmonella typhi* (20 mm), low activity (14mm) against *Pseudomonas aeruginosa* and inactive against *Escherichia coli* were reported by Nishant Rai et al., (2011). The methanol extract of *Hibiscus* has got phytomedical property it may be due to the nature of biologically active compounds present in *hibiscus* whose activity are enhanced in the presence on methanol and also methanol has an stronger extraction capacity which could have produced greater number of active constituents responsible for antibacterial activity (Barker et al., 1995). Plant based antimicrobial compounds have enormous therapeutic potential as they can serve the purpose without any side effects that are often associated with synthetic antimicrobials. The methanol, ethanol, ethyl acetate and chloroform and aqueous extracts of the leaves of *A. asp era, A. parviflora, A. indica*, and *C. odorata* were subjected to a preliminary screening for antimicrobial activity against two human pathogenic bacteria *E. coli* and *S. aureus*. High activity against the Gram-positive organism *E. coli* was found in aqueous and all tested solvent extracts of *A. indica*. In case of human pathogenic *S. aureus*, maximum inhibition of 8 mm was obtained in aqueous extracts of *A. indica*. Similar observations were reported from nimboide isolated from neem seed oil showing antibacterial activity against *S. aureus* and *Staphylococcus coagulase* (Nazma and Rao, 1977). In the present investigation, the antibacterial activity of *Azadirachta indica* plant extract was exhibited maximum zone of inhibition against *Escherichia coli* 14 mm (Mean value in Dia.), when compared with other medicinal plant extracts. *Ficus religiosa* plant extract was zone of inhibition 13 mm (Mean value in Dia.) against *Escherichia coli* and 11 mm (Mean value in Dia.) against *Salmonella typhi, Staphylococcus aureus* and *Enterobacter aerogen*. The *Azadirachta indica* and *Ficus religiosa* medicinal plant extracts were exhibited higher antibacterial activity.

**REFERENCES**


