

ISSN: 2230-9926

### **RESEARCH ARTICLE**

Available online at http://www.journalijdr.com



International Journal of Development Research Vol. 14, Issue, 04, pp. 65529-65532, April, 2024 https://doi.org/10.37118/ijdr.28192.04.2024



**OPEN ACCESS** 

### ANTAGONISTIC EFFECT OF CERAMIC INDUSTRY SOIL STREPTOMYCESAGAINST PLANT PATHOGENS

### \*1Sreeja Bopin and <sup>2</sup>Kalavati Prajapati

<sup>1</sup>Research Scholar, Department of Microbiology, HVHP Institute of Post Graduate Studies and Research, KSV Kadi, (North Gujarat), India; <sup>2</sup>Assistant Professor, Department of Microbiology, Shri PHG Muni. Arts and Sci. College, Kalol (North Gujarat), India

#### **ARTICLE INFO**

#### Article History:

Received 18<sup>th</sup> January, 2024 Received in revised form 20<sup>th</sup> February, 2024 Accepted 19<sup>th</sup> March, 2024 Published online 30<sup>th</sup> April, 2024

#### Key Words:

Streptomycetes, Antagonistic effect, Phytopathogenic organisms, Ceramic soil, potassium solubilizing actinomycetes.

\*Corresponding author: Laxmi Undi,

### ABSTRACT

The various diseases that occur during the growth of plants usually cause a significant reduction in production and quality of agricultural products. Actinomycetes, especially *Streptomyces* spp., become a valuablebiological control resource due to their preponderant abilities to produce various secondary metabolites with novel structure and remarkable biological activity. The potassium solubilizing actinomycetes isolated from the ceramic soil industries were carried out anantagonistic effect against various soil borne phytopathogenic fungiand bacteria. Synthetic bactericides and fungicides have been causing harm to humans, animals, and the environment, as well as generating resistance in phytopathogenic organisms. Actinomycetes produce secondary metabolites with antifungal properties. Approximately 80% of antibiotics, such as streptomycin, spectinomycin, tetracycline and erythromycin, etc., are produced by actinomycetes. Organic acids produced by most of the actinomycetes inhibit the growth of phytopathogenic organisms and promote plant growth and induce systemic resistance in plants. Therefore, the objective of this study is to compare the antagonistic effect of actinomycetes have equal or greater potential as biocontrol agents against phytopathogenic microbes.

Copyright©2024, Sreeja Bopin and Kalavati Prajapati. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Citation: Sreeja Bopin and Kalavati Prajapati, 2024.* "Antagonistic effect of ceramic industry soil streptomycesagainst plant pathogens". International Journal of Development Research, 14, (04), 65529-65532.

# **INTRODUCTION**

Gram-positive aerobic bacteria, or actinomycetes, are widely distributed and exhibit a wide range of morphologies. Some of these bacteria, like those in the Streptomyces genus, have a resemblance to 1997). filamentous fungi (Ronald, These bacteria are ofbiotechnological interest because of their capacity to generate various bioactive chemicals. Actinomycetes have actually been the source of around two thirds of naturally occurring antibiotics (Buckingham, 1997; Newman et al., 2003). The agricultural and pharmaceutical industries find the microorganisms to be a desirable source of natural chemicals. As a result, efforts to screen microorganisms for bioactive compounds (enzymes and secondary metabolites) that are antagonistic to plant diseases have advanced. Numerous studies have examined the uses of microorganism as biocontrol agents in place of chemical fungicides in agriculture (Welbaum et al. 2004; Singh and Chhatpar 2011). Actinomycetes belonging to the Streptomyces genus are highly recognized for their capacity to impede an extensive variety of fungal phytopathogens, making them valuable microorganisms (Gomes et al. 2000). In this current research actinobacteria from soil and healthy plant tissues and screening them for in vitro antagonistic activity against bacterial and

fungal phytopathogens, keeping in mind the significance of actinomycetes in biocontrol and plant growth promotion.

# **MATERIALS AND METHODS**

**Isolation of the Actinomycetes:** Twenty-two strains of potassiumsoluble actinomycetes were recovered from the ceramic industry soil in the Gujarat regions of Kadi, Meshana, and Morbi. Using Khandeparkar's selection ratio, a secondary screening process was conducted on the various isolates to determine their capacity for increased potassium solubilization. Standard culture, morphological, and analysis of the 16S rRNA gene sequence has been performed to select and identify the two actinomycetes strains, *Streptomyces fenghuangensis* (KSA 09) and *Streptomyces atacamensis* (KSA 16).

*Methods Used in the Experiment:* Actinomycetes from the KSA 09 and 16 strains were numbered, single colonies were chosen, purified, and cultivated for five days at room temperature. Using a sterile inoculation loop, pure colonies were obtained through peering at the actinomycete colonies' thin filaments under a light microscope. For further research, the isolated colonies were kept intact.

Selection of Pathogenic Bacteria and Fungi: Pathogenic bacterial strains, including Pseudomonas, Xanthomonas, and Bacillus species, were cultivated every 45 days on petri plates with nutritional agar at 4°C in a laboratory. Pathogenic fungus strains, including Rhizopus and Fusarium species, were cultivated every two months in a laboratory setting using petri plates with Potato dextrose agar (PDA) at 4°C. Subsequently, the promising isolates KSA 09 and 16 strains were introduced into PDA (Potato dextrose agar) medium for fungal cultures at room temperature and NA (Nutrient agar) media for bacterial cultures at 37°C. A subclass of prokaryotic organisms within the phylum Gram-positive bacteria are known as actinomycetes. The majority of them belong to the Actinomycetales order and subclass Actinobacteridae. Filamentous bacteria that belong to this order can create aerial and substrate mycelium, two different forms of branching mycelium. Their DNA has a high G+C concentration (>55 mol%), which sets them apart in part (Stackbrandt et al., 1997). The interactions between rhizosphere actinomycetes that promotes plant growth and elicit defences against plant diseases are brought about by root exudates. Actinomycetes and plants communicate by exchanging nutrients to survive. Based on the amount of nutrients in the soil and the actinomycetes' capacity to communicate with the host plant, several kinds of interactions may form (Hassan et al., 2019).

A rhizospheric actinomycete isolate that was investigated by El-Sayed et al. (2023) had distinct and promising antagonistic activity against three of the most prevalent phytopathogenic fungi: Alternaria brassicicola CBS107, Rhizoctonia solani To18, and Fusarium oxysporum MH105. Based on spore shape and cell wall chemotype, the antagonistic strain was identified, and it was hypothesized that it is a member of the Nocardiopsaceae family. In addition, 16S rRNA gene phylogenetic analysis (OP869859.1) and cultural, physiological, and biochemical traits verified the strain's identity as Nocardiopsis alba. The strain's antifungal efficacy was assessed using its cell-free filtrate (CFF), and the tested fungal species showed inhibition zone widths ranging from  $17.0 \pm 0.92$  to  $19.5 \pm 0.28$  mm. Using a modified version of Dikin et al.'s protocol, the most powerful isolate was further evaluated using the dual culture plate assay against phytopathogenic fungal strains to confirm its antifungal activity (2006). Furthermore, it is one of the primary actinomycetes whose in vitro antifungal activity has been studied to suppress phytopathogenic fungi (Torres-Rodriguez et al., 2022). Examples of these include soil Nocardiopsis dassonvillei, which has been shown to suppress Bipolaris sorokiniana in wheat (Allali et al., 2019), airborne Nocardiopsis alba, which has been shown to suppress Ganoderma boninense (Widada et al., 2021), and soil Nocardiopsis sp., which has been shown to reduce Fusarium sp. (Adlin et al., 2019). Based on chemotaxonomy, phenotypic traits, and 16S rDNA sequence phylogenetic analysis, Streptomyces flaveus was determined to be the strain that suppressed all tested pathogens, SO1. According to Aquar et al. (2020), the results suggest that the SO1 strain has the potential to be used as a biocontrol agent and to boost plant growth. Using partial 16S rRNA sequencing, actinomycetes isolated from the rhizosphere soil were screened by Abdelrahman et al. (2022) for their ability to inhibit the growth of the fungal plant pathogen Phytophthora infestans through metabolic activity. Based on these results, several of the isolates were identified as Nocardiopsis spp.

Identification of Antagonistic Effect of Actinomycete Isolates: The cross-streak approach was utilized to identify the antagonistic effect against specific bacteria and fungi. For additional research, the pure KSA 09 & 16 strains with the strongest antibacterial activity were chosen. The cross-streak method was used to estimate the pathogenic bacteria and fungal inhibitory distance after they were incubated at 37 °C for 24 hours and 25 °C for 48 hours respectively, in order to assess the antimicrobial activity. The cross-streak method (Lemos *et al.*, 1985) was used for primary screening against specific fungi and yeasts, and the agar cylinder method (Disk, 2007) was used for secondary screening against specific pathogenic bacteria (Taechowisan *et al.*, 2005). For additional research, the isolates with the strongest antibacterial activity were chosen.

The formula for calculating the percentage of inhibition was (%) inhibition = (R control- R test)/R control x 100. where R control is the pathogenic fungus's colony diameter on SDA plates and R test is the pathogenic fungus's colony diameter on SDA plates with actinomycete isolates (Wang et al., 2002). The inhibition zone of inhibition's average diameter is used to classify the isolates' level of antibacterial activity. In this instance, the zone of inhibition's diameter was split into four categories: weak activity (≤9 mm), moderate activity (10-12 mm), outstanding activity (≥18 mm), and good activity (12-15 mm). Three separate samples were used. The ability of actinomycetes belonging to the genus Streptomyces to inhibit the growth of numerous fungal pathogens in vitro and in planta is widely recognized (El-Abyad et al., 1993; Abd-Allah, 2001; Coombs et al., 2004). The antagonistic activity of wheat (Triticum aestivum L.) rhizosphere isolates against specific root-rotting fungi (Fusarium graminearum, Fusarium culmorum, Bipolaris sorokiniana and Fusarium verticilloides,) were investigated. These are some of the action modes of the Streptomycetes in the rhizosphere. On solid media, the in vitro antagonistic effects of actinomycetes isolates against fungal pathogens were ascertained. Investigations were done on the mechanism of inhibition, the impact of application time, and pH on inhibition. With respect to all fungi, the actinomycete isolate 129.01 demonstrated a high inhibition ratio of greater than 60%. Under greenhouse circumstances, the isolate 129.01's ability to combat fungi that cause root rot was evaluated. 16S rRNA analysis supported the genus Streptomyces (Erginbaset al., 2010).

## RESULTS

The findings showed that isolates KSA 09 had low antifungal activity but broad-spectrum activity against pathogenic bacteria (Grampositive and Gram-negative), particularly against Pseudomonas spp. (Table 6.1). Most isolates grew best at a temperature of 35 to 37 0 C. Bacillus spp., (11.5 mm), Pseudomonas spp., strain 1 (13 mm), strain 2 (10.3 mm) and Xanthomonas spp., (12 mm) were the bacterial isolates that KSA 09 antagonistically affected, whereas KSA 16 in Bacillus spp., (6.5 mm), Pseudomonas spp. strain 1 &2) (8.5 & 6 mm), and Xanthomonas spp., (8 mm) were the isolates (Table 3. 1 and 3.2). The antagonistic research of KSA 09 and 16 against pathogenic fungal strains, including Fusarium spp. and Rhizopus spp., revealed that the former was less sensitive to KSA 09 than the latter to KSA 16. This finding verified that KSA 09, which is highly recommended, has an antagonistic effect on phytopathogens.

 
 Table 1. Antagonistic Effect of Fungal Cultures with Zone of Inhibition

Fungi cultures	Zone of Inhibition (mm)		
	KSA 09	KSA 09	
Aspergillus terreus	10.5	8	
Rhizopus spp.	8.5	6.5	

Table 2. Antagonistic Effect of Bacterial Cultures with Interpretation

Bacterial cultures	Zone of Inhibition (mm)	
	KSA 09	KSA 16
Xanthomonas spp.,	12	8
Pseudomonas spp. (St.1)	13	8.5
Pseudomonas spp. (St.2)	10.3	6
Bacillus spp.	11.5	6.5

Table . 3Antagonistic effect of Fungi cultures with Zone of Inhibition

Bacterial cultures	KSA 09	KSA 16
Xanthomonas spp.,	Moderate sensitive	Less sensitive
Pseudomonas spp. (St.1)	Highly sensitive	Less sensitive
Pseudomonas spp. (St.2)	Moderate sensitive	Less sensitive
Bacillus spp.	Moderate sensitive	Less sensitive

#### Table 4. Antagonistic effect of Fungi cultures with Interpretation

Fungi cultures	KSA 09	KSA 16
Aspergillus terreus	Moderate sensitive	Less sensitive
Rhizopus spp.	Moderate sensitive	Less sensitive

Actinomycetes, particularly Streptomyces, have been a subject of substantial study interest over the past 60 years due to their ability to create several physiologically active chemicals as part of this Grampositive bacterial group. According to Silhavy et al. (2010), the isolates' crude extracts demonstrated antibacterial activity against Gram-negative bacteria, which are frequently more resistant to antimicrobial treatments than Gram-positive bacteria. Actinobacteria's antagonistic effects on pathogenic fungus have been evaluated across a wide range of fungal species. Various studies have demonstrated the capability of actinobacteria isolates to counteract phytopathogenic fungi (Aouar et al. 2012; Goudjalet al. 2016; Abbasi et al. 2019). Putrieet al. (2020) identified actinobacteria as the predominant microorganisms in the rhizospheric soil. Khamna et al. (2009) showed that microorganisms from the rhizosphere are best for controlling plant diseases, which is why we have studied rhizospheric soil. Actinomycete isolates were isolated and screened by Kaur et al. (2013) for potential antagonistic activity and activities that promote plant growth. Of the 321 isolates, 156 were rhizospheric, 103 were non-rhizospheric, and 62 were endophytic. Eighty-three isolates demonstrated antagonistic activity against one or more test phytopathogenic fungi during the dual culture assay primary screening. Out of the activeisolates, 20 showed antagonistic properties in the well diffusion experiment during the secondary screening. The majority of them exhibited a wide-ranging inhibitory effect against five to six test fungi. The study's findings suggest that these isolates have potential applications as biocontrol and plant growth boosting agents. Streptomyces was the genus that all of the active isolates belonged to, according to morphological and chemotaxonomic investigations. Considering the aforementioned results, the objective was to isolate actinomycetes with potent antibacterial properties from ceramic soils, which serve as a model for a harsh ecology. Out of the 22 actinomycetes found in ceramic soils, Streptomyces fenghuangensis (KSA 09) and Streptomyces atacamensis (KSA 16) showed wide-ranging phytopathogenic effects against various harmful bacteria and fungi. The KSA 09 and KSA 16 isolates were identified as Streptomyces fenghuangensis and Streptomyces atacamensis by a combination of conventional and molecular approaches.

## CONCLUSION

An additional round of antimicrobial screening was performed on each isolate to check for harmful fungus and bacteria. According to the results, two of the isolates (KSA 09 and KSA 16) exhibited antagonistic effect against the majority of the pathogenic bacteria that were tested. Consequently, these two intriguing isolates were chosen for further identification after being previously determined to be Streptomyces via morphological, biochemical, and physiological techniques. The findings showed that isolate KSA 09 had modest antifungal activity but broad-spectrum action against pathogenic bacteria (Gram-positive and Gram-negative), particularly against Pseudomonas spp. Most isolates grew best at a temperature of 25 to 30 degrees Celsius. These results validated KSA 09's antagonistic action against phytopathogens, which is highly recommended.

Acknowledgement: The author is thankful to the Department of Microbiology, HVHPGR, Kadi and Management of Kadi Sarva Vishwavidyalaya (KSV), Gandhinagar, for providing the facilities for this research.

## REFERENCES

Abbasi S, Safaie N, Sadeghi A, Shamsbakhsh M. 2019. Streptomyces strains induce resistance to Fusarium oxysporum f. sp. lycopersici race 3 in tomato through different molecular mechanisms. *Front Microbial*. DOI: 10.3389/fmicb.2019.01505.

- Abd-Allah, E. F. (2001) Streptomyces plicatus as a model biocontrol agent, *Folia Microbiol.* 46, 309-314.
- Abdelrahman O, Yagi S, El Siddig M, El Hussein A, Germanier F, De Vrieze M, et al. 2022. Evaluating the antagonistic potential of actinomycete strains isolated from Sudan's soils against Phytophthora infestans. *Front. Microbiol.* 13: 82782
- Adlin Jenifer JSC, Michaelbabu M, Eswaramoorthy Thirumalaikumar CL, Jeraldin Nisha SR, Uma G, Citarasu T. 2019. Antimicrobial potential of haloalkaliphilic Nocardiopsis sp. AJ1 isolated from solar salterns in India. J. Basic Microbiol. 59: 288-301.
- Allali K, Goudjal Y, Zamoum M, Bouznada K, Sabaou N, Zitouni A. 2019. Nocardiopsis dassonvillei strain MB22 from the Algerian Sahara promotes wheat seedlings growth and potentially controls the common root rot pathogen Bipolaris sorokiniana. J. Plant Pathol. 101: 1115-1125.
- Aouar L, Lerat S, Ouffroukh A, Boulahrouf A, Beaulieu C. 2012. Taxonomic identification of rhizospheric actinobacteria isolated from Algerian semi-arid soil exhibiting antagonistic activities against plant fungal pathogens. *Can J Plant Pathol* 34 (2): 165-176. DOI: 10.1080/07060661.2012.681396.
- Aouar, L., Boukelloul, I., & Benadjila, A. (2020). Identification of antagonistic Streptomyces strains isolated from Algerian Saharan soils and their plant growth promoting properties. *Biodiversitas Journal of Biological Diversity*, 21(12).
- Audrey W. Disk Diffusion Test and Gradient Methodologies. In: Schwalbe R, Steele-Moore L, Goodwin AC, editors. Antimicrobial Susceptibility Testing Protocols: Boca Raton: CRC Press Taylor and Francis Group; 2007. p. 53-72.
- Barriuso, J., Ramos Solano, B., and Gutierrez Manero, F. J. (2008). Protection against pathogen and salt stress by four plant growthpromoting rhizobacteria isolated from Pinus sp. on Arabidopsis thaliana. *Biological Control*, 98: 666–672.
- Boudemagh A, Kitouni M, Boughachiche F, Hamdiken H et al. 2005. Isolation and molecular identification of actinomycetes microflora, of some Saharan soils of south east Algeria (Biskra, El-Oued and Ourgla) study of antifungal activity of isolated strains. J Med Mycol 15: 39-44. DOI: 10.1016/ j.mycmed.2004.12.004.
- Buckingham J. Dictionary of Natural Products, Supplement 4: Taylor & Francis; 1997.
- Coombs, J.T., Michelsen P.P. and Franco, C.M.M. (2004) Evaluation of endophitic actinobacteria as antagonists of Gaeumannomycesgraminis var. tritici in wheat. Biol. Control. 29, 359-366.
- Dikin A, Sijam K, Kadir J, Semanz IA. 2006. Antagonistic bacteria against Schizophyllum commune Fr. in Peninsular Malaysia. Biotropia 13: 111-121
- El-Abyad, M. S., El-Sayed, M. A., El-Shanshoury A. R. and El-Sabbagh, S.M. (1993) Towards the biological control of fungal and bacterial diseases of tomato using antagonistic Streptomyces spp. *Plant Soil.* 149, 185-195.
- Erginbas, G., Yamac, M., Amoroso, M. J. D. R., & Cuozzo, S. A. (2010). Selection of antagonistic actinomycete isolates as biocontrol agents against root-rot fungi, *Fresenius Environmental Bulletin*, 19(3), 417-424.
- Gomes RC, Semedo LT, Soares RM, Linhares LF, Alviano CS, Coelho RR. 2000. Chitinolytic activity of Actinomycetes from a cerrado soil and their potential in biocontrol. *Lett Appl Microbiol.* 30:146–150.
- Goudjal Y, Zamoum M, Meklat A, Sabaou N, Mathieu F, Zitouni A. 2016. Plant-growth-promoting potential of endosymbiotic actinobacteria isolated from sand truffles (Terfezialeonis Tul.) of the Algerian Sahara. *Ann Microbiol* 66 (1): 91-100. DOI: 10.1007/s13213-015-1085-2.
- Hassan, M. K., McInroy, J. A., Kloepper, J. W. (2019). The Interactions of Rhizodeposits with Plant Growth-Promoting Rhizobacteria in the Rhizosphere: A Review. Agriculture, 9 (142): 1-13.
- Jeon, J. S., Lee, S. S., Kim, H. Y., Ahn, T. S., and Song, H. G. (2003). Plant growth promotion in soil by some inoculated microorganisms. *Journal of Microbiology*, 41: 271–276.

- Kaur, T., Sharma, D., Kaur, A., & Manhas, R. K. (2013). Antagonistic and plant growth promoting activities of endophytic and soil actinomycetes. *Archives of Phytopathology and Plant Protection*, 46(14), 1756-1768.
- Khamna S, Yokot A, Lumyong S. 2009. Actinomycetes isolated from medicinal plant rhizosphere soils: diversity and screening of antifungal compounds, indole-3-acetic acid and siderophore production. *World J Microbiol Biotechnol* 25: 649-655. DOI: 10.1007/s11274-008-9933-x.
- Khan, A. G. (2005). Role of soil microbes in the rhizospheres of plants growing on trace metal contaminated soils in phytoremediation. *Journal of Trace Elements in Medicine and Biology*, 18: 355–364.
- Lemos ML, Toranzo AE, Barja JL. Antibiotic activity of epiphytic bacteria isolated from intertidal seaweeds. *Microb Ecol* 1985;11:149-63.
- Lucy, M., Reed, E., and Glick, B. R. (2004). Application of free living plant-promoting rhizobacteria. Antonie van Leeuwenhoek, 86: 1–25.
- Majeed, A., Abbasi, M. K., Hameed, S., Imran, A. and Rahim, N. (2015). Isolation and characterization of plant growth-promoting rhizobacteria from wheat rhizosphere and their effect on plant growth promotion. Frontiers in Microbiology, 6: 198.
- Marques, A. P. G. C., Pires, C., Moreira, H., Rangel, A. O. S. S., Castro, P. M. L. (2010). Assessment of the plant growth promotion abilities of six bacterial isolates using Zea mays as indicator plant. Soil Biology and Biochemistry, 42: 1229–1235.
- Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981-2002. J Nat Prod 2003;66:1022-37
- Nimnoi P, Pongsilp N, Lumyong S. 2010. Endophytic actinomycetes isolated from Aquilaria crassna Pierre ex Lec and screening of plant growth promoters production. *World J Microb Biotechnol.* 26:193–203.

- Putrie RFW, Aryantha INP, Iriawati I, Antonius S. 2020. Diversity of endophytic and rhizosphere bacteria from pineapple (Ananas comosus) plant in semi-arid ecosystem. Biodiversitas 21 (7): 3084-3093. DOI: 10.13057/biodiv/d210728
- Ronald M. Principles of microbiology. New York: WCB McGrill-Hill; 1997.
- Silhavy TJ, Kahne D, Walker S. The Bacterial Cell Envelope. Cold Spring Harbor Perspectives in Biology 2010;2:a000414.
- Singh AK, Chhatpar HS. 2011. Combined use of Streptomyces sp. A6 and chemical fungicides against Fusarium wilt of Cajanus cajan may reduce the dosage of fungicides used in the field. Crop Prot. 30:770–775.
- Stackbrandt, E., Rainey, F. A. and Ward Rainey, N. L. (1997). proposal for new hierarchic classification system ,Actinobacteria classic nov. International Journal of Systematic Bacteriology, 47: 479-491.
- Taechowisan T, Lu C, Shen Y, et al. Secondary metabolites from endophytic Streptomyces aureofaciens CMUAc130 and their antifungal activity. Microbiology 2005;151:1691-5.
- Torres-Rodriguez JA, Reyes-Pérez JJ, Quiñones-Aguilar EE, Hernandez-Montiel LG. 2022. Actinomycete potential as biocontrol agent of phytopathogenic fungi: mechanisms, source, and applications. Plants (Basel) 11: 3201.
- Wang SL, Hsiao WJ, Chang WT. Purification and characterization of an antimicrobial chitinase extracellularly produced by Monascuspurpureus CCRC31499 in a shrimp and crab shell powder medium. J Agric Food Chem 2002;50:2249- 55.
- Welbaum G, Sturz AV, Dong Z, Nowak J. 2004. Fertilizing soil microorganisms to improve productivity of agroecosystems. Crit Rev Plant Sci. 23:175–197.
- Widada J, Damayanti E, Alhakim MR, Yuwono T, Mustofa M. 2021. Two strains of airborne Nocardiopsis alba producing different volatile organic compounds (VOCs) as biofungicide for Ganoderma boninense. *FEMS Microbiol.* Lett. 368: fnab138.

\*\*\*\*\*\*