



ISSN: 2230-9926

Available online at <http://www.journalijdr.com>

IJDR

International Journal of Development Research
Vol. 10, Issue, 03, pp. 34243-34247, March, 2020



RESEARCH ARTICLE

OPEN ACCESS

PHYSICOCHEMICAL CHARACTERISTICS OF RED PITAYA IN TWO REGIONS OF THE MATO GROSSO DO SUL STATE, BRAZIL

¹Suni Liu, ²Jade Oliveira Santos, ^{*3}Verônica Assalin Zorgetto-Pinheiro, ³Maria Margarida Morena Domingos Levenhagen, ⁴Liu Hsuan Han, ⁵Rita de Cássia Avellaneda Guimarães, ⁵Raquel Pires Campos, ⁵Danielle Bogo and ⁵Luciana Miyagusku

¹Master's Degree Fellow, Health and Development of the West-central Region Graduate Program, Federal University of Mato Grosso do Sul, Campo Grande-MS, Brazil; ²Bachelor of Nutrition, Federal University of Mato Grosso do Sul, Campo Grande-MS, Brazil; ³PhD Fellow, Health and Development of the West-central Region Graduate Program, Federal University of Mato Grosso do Sul, Campo Grande-MS, Brazil; ⁴PhD Fellow, Chemistry Graduate Program, Federal University of Mato Grosso do Sul, Campo Grande-MS, Brazil; ⁵PhD, Adjunct Professor, Federal University of Mato Grosso do Sul, Campo Grande-MS, Brazil

ARTICLE INFO

Article History:

Received 19th December, 2019
Received in revised form
03rd January, 2020
Accepted 17th February, 2020
Published online 30th March, 2020

Key Words:

Hylocereus spp.; Cactaceae; functional food; red pitaya; physicochemical characteristics.

*Corresponding author: Verônica Assalin Zorgetto-Pinheiro,

ABSTRACT

The pink pitaya with red pulp (*Hylocereus* spp.) is spreading in the Brazilian markets for its exotic characteristics and nutritional value; however, there is still not enough reports about its postharvest characteristics. The aim of this research was to characterize physical and chemical features and nutritional values of postharvest pitayas. The fruits were collected in two different regions of Mato Grosso do Sul State, Brazil, and these samples were subjected to the following evaluations: length, diameter, fresh matter, moisture content, fixed mineral residue; soluble solids, pH, titratable acidity, carbohydrate, protein, lipids, total phenolic compounds, tannins; and total antioxidants. Pitaya has low lipid and protein levels, with high water contents (86-89%) and carbohydrates being the highest macronutrient value present in the fruit. The soluble solids content (^oBrix 14,9) and the low titratable acidity suggests that the fruit is a good option of *in natura* consumption. Bioactives were more present in the pulp than in the peel, except in tannins contents. Pitaya can be classified as a functional food, thus acting in the prevention of diseases, because it contains compounds that fight free radicals.

Copyright © 2020, Suni Liu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Suni Liu, Jade Oliveira Santos, Verônica Assalin Zorgetto-Pinheiro, Maria Margarida Morena Domingos Levenhagen et al. 2020. "Physicochemical characteristics of red pitaya in two regions of the Mato Grosso do Sul State, Brazil", *International Journal of Development Research*, 10, (03), 34243-34247.

INTRODUCTION

Pitaya (*Hylocereus* spp.) is an exotic fruit, originated from America, that belongs to the Cactaceae family. In Brazil, the growing demand for exotic fruits and their high added value arouse the interest of commercialization of Pitaya, with the Southeast region being the major producer (Bastos et al., 2006). Between 2007 and 2012 there was an increase in production from 81 to 299 tons / year, corresponding to more than 250% in the marketing of pitaya at the Company of General Warehouses of the State of São Paulo (CEAGESP) with greater demand from December to April (Watanabe and Oliveira, 2014). Because of its appearance and shape is commonly known as dragon fruit. It is found in countries such as Costa Rica, Venezuela, Panama, Uruguay, Brazil, Colombia

and Mexico, being the two-last cited, the largest world producers (Canto et al., 1993). Currently, the main commercial Pitaya species are the white-fleshed red pitaya (*Hylocereus undatus* (Haw.) Britton & Rose) and the white-fleshed yellow pitaya (*Selenicereus megalanthus* (K Schum ex Vaupel) (Nerd et al., 2002). Pitaya is harvested between December to April and its fruits develop at temperatures between 18°C and 26°C. Although its species are usually cultivated in a dry tropical climate (Junqueira et al., 2002) and are influenced directly by rainy periods, they can adapt to different types of climates (Donadio, 2009). There is a diversity of characteristics between the species such as: shape, presence of thorns, color of the peel and the pulp, as well as its pH and soluble solids content. In general, pitaya has a sweet and mild flavor, a firm pulp, being sources of vitamins and minerals, attracting consumer interest (Cordeiro et al., 2015).

Red Pitaya is pointed out as a functional food due to the presence of phenolic compounds, mainly betalains, that have considerable antioxidant activity, and also can be used as natural source for food pigments. Thus, being associated with the prevention of diseases such as cancer and to reduced risk of mortality from cardiovascular diseases (Stintzing *et al.*, 2004; Vaillant *et al.*, 2005; Herbach *et al.*, 2006; Wu *et al.*, 2006). In addition to the betalains, the Pitaya pulp is also composed by other antioxidants such as flavonoids (Stintzing *et al.*, 2004), oligosaccharides with prebiotic properties (Escribano *et al.*, 2002), soluble fibers that can assist the digestive process, also associated with the regulation of sugar blood levels in patients with type 2 diabetes (Zainoldin and Baba, 2009). The seeds are small and soft, and well distributed by the pulp, constituting 1-2% of the fruit. The pitaya seeds oil is rich in essential fatty acids, especially linoleic acid, that can present a mild laxative function (Crane and Balerdi, 2005), also contributing to the decrease of LDL cholesterol (low density lipoprotein), by inhibiting the absorption of cholesterol in the intestine (Sato *et al.*, 2014). Although the Pitaya consumption has been growing worldwide, there is still a lack of sufficient database of the chemical composition of this potentially nutritive fruit, especially regarding the red pitaya (*Hylocereus spp.*). Based on this, the objective of this study was to evaluate the physical and chemical differences of the red pitaya pulp and peel between two different regions in the Mato Grosso do Sul State.

MATERIALS AND METHODS

Collection of the fruits: Pitaya's fruits were collected in two different regions of the Mato Grosso do Sul State, in the cities of Campo Grande and Ponta Porã. The chemical analyzes were carried out at the Food Technology and Public Health Unit (UTASP) located at the Federal University of Mato Grosso do Sul (UFMS) in Campo Grande, MS. One kilogram of fruits was collected from each location, properly cleaned with a 1% hypochlorite solution for 15 minutes, and subsequently rinsed under water to eliminate chlorine residues. Physical measurements were performed, and diameter, weight of the total fruit mass, weight of the pulp and peel were obtained.

Longitudinal and transversal diameter and fresh weight: The total mass of the fruit, peel, pulp, total length and total diameter were determined using the methodology of Lima (2013). The fruits were weighted on a semi-analytical scale, and the diameter was measured with the aid of a pachymeter.

Titrateable acidity, soluble solids and pH: The soluble solids content was determined with the aid of a portable refractometer (Brasil, 2005). The total titrateable acidity was performed in the pulp and peel of the crushed and homogenized fruit by titration with 0.10 M NaOH solution until a pH 8.1. This experiment was performed with constant monitorization of a pHmeter and the results expressed in g / 100g of citric acid (% m / v).

Centesimal composition and caloric value of the fruit: The compounds of moisture, protein, carbohydrates, lipids and fixed mineral residue were determined in the pulp and peel crushed and homogenized according to the methodologies of the Adolfo Lutz Institute (Brasil, 2005) and AOAC (2002). The carbohydrate content was measured by difference. The caloric value was calculated by the protein, carbohydrate and

lipid content of the samples, using the Atwater factors, described by Mahan and Escott-Stumo(2005). The results were expressed in kilocalories (kcal), according to the following equation:

$$\text{Calories value (kcal)} = (\text{protein content} \times 4.0) + (\text{carbohydrate content} \times 4.0) + (\text{lipid content} \times 9.0)$$

$$\% \text{ Carbohydrates} = 100 - (\% \text{ moisture} + \% \text{ fixed mineral residue} + \% \text{ proteins} + \% \text{ lipids})$$

Antioxidant activity: The antioxidant activity was determined by spectrophotometry in the visible region ($\lambda = 517 \text{ nm}$) using the DPPH radical sequestration method (2,2 diphenyl-1-picrilhidrazil) with modifications from Rufino *et al.*(2007). The results were expressed using the following equation, where % SRL = percentage of free radical scavenging; Ac = absorbance of the control; Ae = absorbance of the extract:

$$\% \text{ SRL} = (\text{Ac} - \text{Ae}) \times 100 / \text{Ac}$$

Total phenolic compounds: The phenolic compounds were dosed with spectrophotometry in the visible region ($\lambda = 760 \text{ nm}$) as proposed by Waterhouse (2002), using the Folin-Ciocalteu reagent. The results were expressed in mg of gallic acid equivalent (GAE) per 100g of pulp or peel.

Tannins: The tannins were extracted and its quantification was performed by the colorimetric method of Folin- Denis n° 952.03 from the AOAC (2002). The results were expressed in mg of tannic acid equivalent (TAE) per 100g of pulp or peel.

Statistical Analysis: The physical-chemical data obtained was evaluated using the analysis of variance (ANOVA), the Tukey test and student's t test in order to compare means.

RESULTS AND DISCUSSION

Fruits total mass average had a variation of 100g between the investigated locations. Ponta Porã fruits obtained the great variation in the weight, achieving the minimum of 232.95g and the maximum of 357.78g. The parameters (length and height) were similar between them, as shown in Table 1. The fresh mass of pitaya fruits found in our study was similar to some previous researches of other groups, that described an average of $411.22 \text{ g} \pm 26.17$ for the fresh mass (Cordeiro *et al.*, 2015).

Table 1. Physical characteristics of *Hylocereus spp* fruits from two different regions of the Mato Grosso do Sul State

Parameters	Campo Grande	Ponta Porã
Freshmass (g)	416,77 ±28,73	305,44 ±119,83
Length (cm)	28,40 ±0,2	26,23 ±2,0
Height (cm)	30,20 ±2,0	27,50 ±4,1
Peel (%)	16,40	16,63
Pulp (%)	77,86	76,66

Other research, that analyzed fruits from three different cities in the Pará State, also in Brazil, found values between 351.25g and 430g for the total weight of the fruit, indicating a great variation between locations. However, they found no significant variation in the pulp percentage, and other measurements, even for fruits grown with different types of fertilizer (Sato *et al.*, 2014). However, the parameters of the present study were higher when compared to other authors, with a length of 8.59 - 10.7 cm and diameter of 7.77 - 8.70 cm (Pinto *et al.*, 2010; Takata, 2012; Sato *et al.*, 2014; Cordeiro *et al.*, 2015), while in this study, the values were 25 - 28.3 cm for

length and 25 - 31.1 cm in diameter for both regions studied. Pitaya pulp accounts for about 60 to 80% of the fruit weight, and consists of 82-88% of water according to Oliveira *et al.* (2010). The values found in the present work were approximately 77% of the fruit weight for the pulp and 16% for the peel (Table 1).

Protein content was higher in pulp than in the peel, corresponding to 1.14-1.69% and 0.80-0.83%, respectively. These values were similar to the mean values of 1.12% found by Sato *et al.* (2014), and slightly higher than the value of 1.06% found by Oliveira *et al.* (2010). The pulp obtained higher lipid levels than the peel, 0.54 and 0.56% for the pulp

Table 2. Chemical characteristics of *Hylocereus spp.* fruits from two different regions of the Mato Grosso do Sul State

Parameters	Campo Grande		Ponta Porã	
	Pulp	Peel	Pulp	Peel
Soluble Solids (°Brix)	14,9 ±0,0	5,5 ±0,0	14,9 ±0,0	5,3 ±0,0
Titrateable Acidity*	0,26 ±0,01	0,18 ±0,01	0,27 ±0,01	0,26 ±0,01
pH	4,5 ±0,0	4,6 ±0,0	4,7 ±0,0	4,3 ±0,0

*g/ 100g citric acid

Table 3. Centesimal composition and caloric value of *Hylocereus spp.* fruits from two different regions of the Mato Grosso do Sul State

Parameters	Campo Grande		Ponta Porã	
	Pulp	Peel	Pulp	Peel
Moisture (%)	87,64 ± 0,87	89,99 ±0,31	86,91 ±0,63	89,81 ±0,37
Fixed Mineral Residue (%)	0,67 ±0,07	1,62 ±0,25	0,73 ±0,05	2,00 ±0,05
Proteins (%)	1,14 ±0,43	0,83 ±0,14	1,69 ±0,0	0,80 ±0,05
Carbohydrates (%)	10,01 ±0,0	7,53 ±0,0	10,12 ±0,0	7,09 ±0,0
Lipids (%)	0,54 ±0,03	0,17 ±0,05	0,56 ±0,26	0,20 ±0,08
Antioxidants (IC50 mg/mL)	297,85 ±0,0	283,21 ±0,0	380,71 ±0,0	169,17 ±0,0
Phenols*	16,58 ±0,0	12,77 ±0,0	30,97 ±0,0	14,35 ±0,0
Tannins**	5,29 ±0,0	13,87 ±0,0	12,31 ±0,0	24,37 ±0,0
Caloric Value (Kcal)	49,42		52,28	

*mg GAE/ 100 g⁻¹

**mg TAE/ 100 g⁻¹

The pulp was in a higher proportion than found by Jamilah *et al.* (2011), corresponding to 65% and 22% for peel. Humidity was 86-87% for the pulp, and slightly higher for the peel, 89% (Table 3). The soluble solids content was higher in the pulp, as well as titrateable acidity. The median part is the sweetest of the fruit. The values found for pH are similar between peel and pulp (Lima, 2013). The soluble solids values were higher than that found by Oliveira *et al.* (2010), which obtained 11.00°Brix for pulp, and similar to that obtained by Cordeiro *et al.* (2015), of 13.14°Brix. The present study found a value of 0.26 in titrateable acidity (g. 100g⁻¹), and pH 4.88 (Table 2). Having the tendency of low acidity, pitaya fruits require post-harvest care, since foods with pH higher than 4.5 may have a higher chance of microbial proliferation, both pathogenic and deteriorating strains, while in an acid medium, that is pH ranging from 4.0 to 4.5, there is a higher presence of yeasts, molds and some specific bacterial strains such as the lactic ones (Franco and Landgraf, 2003). Soluble solids consist mainly of glucose and fructose and the higher this value the better the acceptability of *in natura* consumption of these fruits, also related to their low sugar values and little acidity (Stintzing and Schieber, 2003; Vaillant *et al.*, 2005). According to Wall and Khan (2008), sucrose corresponds to 2% of total sugar from pitaya fruit while Esquivel, *et al.* (2007), found sucrose values lower than 1%. Some authors point out that white pitaya has a higher soluble solids content than red pitaya, and that the distribution in the fruit is not homogeneous, being the central part richer in sugars than the extremities. When ripe, fruits generally present higher values of soluble solids, which are advantageous for the agro-industrial sector, aiming at reducing sugar addition during the process (Chitarra and Chitarra, 2005). However, low soluble solids levels require a greater potential for post-harvest conservation, as excess sugars may be associated with faster decay time (Brunini *et al.*, 2003).

and 0.17 and 0.20% for the peel from Campo Grande and Ponta Porã samples, respectively. Pitaya fruits have low lipid content (Pimentel *et al.*, 2005). Sato and co-workers (2014) report lipid values of 0.18 - 0.21% with similar values among three regions studied by them, and are also lower than in other studies, where a value of 0.36% of lipids was found (Oliveira *et al.*, 2010). Such values are mainly provided from pitaya seed, which are numerous and well spread in the mucilaginous fruit pulp. These seeds are rich in essential fatty acids, especially linoleic acid, and can be found in a higher amount than other foods sources, such as flaxseed and canola (Crane and Balerdi, 2005)]. Oliveira *et al.* (2010) concluded that pitaya pulps are rich in carbohydrates and peels are rich in fibers. The total carbohydrates found in the present study were 10.01-10.12% for pulp and 7.53 -7.09% for peel on samples from Campo Grande and Ponta Porã, respectively. These results were slightly lower than presented by other authors, ranging from 10.93 to 12.34% of total carbohydrates (Santo *et al.*, 2014; Oliveira *et al.*, 2010).

Cordeiro *et al.* (2015) obtained a high crude fiber content in their research, of 11.35g in 100g of sample., being considered a fruit of high crude fiber content. Sato *et al.* (2014) concluded in their experiments that pitayas can be considered natural sources of insoluble fibers, finding values of up to 2%. Such results are higher in amount of fiber than fruits such as papaya and cajá (Carvalho *et al.*, 2011). Proper fiber intake has been related to benefits on the digestive tract, among them, action on reducing glucose absorption and combating cardiovascular diseases, obesity and colon diseases (Pimentel *et al.*, 2005). The Food and Drug Administration (FDA) recommends a daily total fiber consumption of 25g based on a 2,000-calorie diet (Cordeiro *et al.* 2015). Sato *et al.* (2014) states that pitaya has a low energy value of about 52 Kcal/100 g, which allows the inclusion of the fruit for healthy eating. Pitayas from Campo Grande presented 49.42Kcal/ 100g, and those from

Ponta Porã, 52.28Kcal/ 100g. This number of calories is similar to fruits like kiwi, red guava and pineapple. The red pitaya has significant mineral contents, especially in relation to calcium, since the high content of this mineral may indicate a higher resistance of the fruit after harvest (Cordeiro *et al.* 2015), because it has a direct relationship between softening, firmness and shelf life (Natale *et al.*, 2005). The present study found a total phenol content higher in the pulp than in the peel. However, some studies have found high levels of phenolic compounds for red pitaya peel, ranging from 40.68mg GAE.100g⁻¹(Mello *et al.*, 2015) to 39.7 mg GAE.100g⁻¹(Wu *et al.*, 2006). The difference between the results of these two regions is probably due to climate variation, and the ripeness state of fruits (Lima, 2013). The values found were considerably lower when compared to other fruits such as orange (75 mg GAE. 100g⁻¹), guava (138 mg GAE. 100g⁻¹) and banana (51 mg GAE. 100g⁻¹). Ascorbic acid present in pitaya contributes significantly to antioxidant activity, as well as acts synergistically with the phenolic capacity to combat the action of free radicals (Cho and Yong, 2011).

Tannins are characterized as phenolic compounds of high molecular mass, which have the ability to precipitate proteins (Efraim and Tucci, 2006). Compounds with antioxidant activity play an important role in preventing oxidative diseases, but in large quantities these same compounds may present undesirable characteristics, such as enzymatic darkening of fruits, interaction with proteins, carbohydrates and minerals (Imeh and Khokhar, 2002). The peel has a greater amount of tannins than compared to the pulp, however, such quantities are not high enough to add undesirable factors to the fruit. No studies were found to compare tannin values for pitaya. Some authors indicate that using the DPPH method for antioxidant analyses in pitayas present a lower activity of this compound. Some authors noted that pitaya peel had higher antioxidant activity when analyzed by a stable radical reduction method, DPPH (177.14µmol AEAC. 100g⁻¹) than by the iron reducing method, FRAP (109.29µmol AEAC. 100g⁻¹) (Abreu *et al.*, 2012). The pulp values found in the literature were 306.81µmol AEAC. 100g⁻¹(Muñoz *et al.*, 2002), similar to the present study for both pulp and peel, with the exception of Ponta Porã peel (IC₅₀:169,0µmol DPPH). The higher the phenolic compound content, the greater the antioxidant action (Wu and Ng, 2008). Antioxidant activity decreases according to fruit ripening time, which is due to the reduction in vitamin C levels and phenolic compounds, since these compounds act as antioxidant factors (Santos *et al.*, 2016).

Betalains are also the main responsible for the antioxidant capacity of red pulp pitaya (Esquivel *et al.*, 2007), having a positive correlation where the higher betalain content, the greater the antioxidant capacity of this fruit (Vaillant *et al.*, 2005). Pitaya peels can be used for natural pigment extraction due to the high presence of betalains, being a good alternative to use of the whole fruit by food industry. In addition, betalains incorporated into blood cells can protect them from oxidative hemolysis and also act inducing quinone reductase, an enzyme associated with cancer chemoprevention(Mello *et al.*, 2015). The variety of genotypes within a pitaya species and consequently different nutrient content in the fruit may have an environmental component, considering that not all fruits analyzed in studies were produced at the same site. Fruits present quantitative and qualitative variations regarding their physicochemical characteristics, due to intrinsic factors (cropping, variety, ripeness stage) and extrinsic (climatic and

edaphic conditions) (Reynerston *et al.*, 2008). Plant coloration is based on some factors such as pigment structure and concentration, pH, temperature, local light intensity, sugars, among others (Stintzing and Carle, 2004). Thus, it is concluded that red pitaya characteristics vary chemically, physically and nutritionally according to the region. Pitaya has characteristics that suggest greater acceptance for *in natura* consumption, highlighting the low acidity and soluble solids content. In addition, almost 90% of the fruit is composed of water, has low concentration of proteins, lipids and the most outstanding content of macronutrients is carbohydrates. The antioxidant compounds present in the fruit can make it act and be classified as a functional fruit, acting in reducing the risk of chronic noncommunicable diseases.

Acknowledgements

This research was financially supported by the Higher Education Personnel Improvement Coordination (CAPES) – Financial Code 001, and by the Federal University of Mato Grosso do Sul, Campo Grande-MS, Brazil.

Conflict of Interest: The authors declare no conflict of interest.

REFERENCES

- Abreu WC, Lopes CO, Pinto KM, Oliveira LA, Carvalho GBM, and Barcelo MFP. 2012. Características físico-químicas e atividade total de pitaia vermelha e branca. *Revista Instituto Adolfo Lutz*. 71 (4): 656-661.
- Association Of Official Analytical Chemists - AOAC. 2002. Official methods of analysis of analysis. 17^a ed. Washington, D.C.
- Bastos DC, Pio, R, Filho JAS, Libardi MN, Almeida LFP, Galuchi TPD, and Bakker ST. 2006. Propagação da pitaya ‘vermelha’ por estaquia. *Ciênc. Agrotec*.30(6):1106-1109.
- Brasil. Agência Nacional de Vigilância Sanitária. 2005. Métodos físico-químicos para análises de alimentos. Brasília: Ministério da Saúde, 4^a ed. Brasília, 1018pp.
- Brunini MA, Oliveira AL, and Varanda DB. 2003. Avaliação da qualidade de polpa de goiaba “Paluma” armazenada a -20°C. *Revista Brasileira de Fruticultura*. 25(3):394-396.
- Canto AR, Albarado JCG, Santarosa MGG, Ramos CJ, García MCM, Hernández LJP, Lazo VR; Medina LR, Rodríguez RR, Torres ET, García SV, and Eloísa EZ. 1993. El cultivo de pitahaya en Yucatán. Gobierno del Estado de Yucatan, Universidad Autónoma Chapingo: México. 53 pp.
- Carvalho AV, Mattietto RA, Assis GT, and Lourenço LFH. 2011. Avaliação do efeito da combinação de pectina, gelatina e alginato de sódio sobre as características de gel de fruta estruturada a partir de “mix” de polpa de cajá e mamão, por meio da metodologia de superfície de resposta. *Acta Amazonica*. 41(2):267-274.
- Chitarra MIF, and Chitarra AB. 2005 Pós-colheita de frutas e hortaliças: fisiologia e manuseio. 2 ed. UFLA: Lavras. Brasil. 785 pp.
- Cho WS, and Yong WK. 2011. Antioxidant properties of two species of *Hylocereus* fruits. *Advanced in Applied Science Research*. 2(3):418-425.
- Cordeiro MHM, Silva JM, Mizobutsi, GP, Mizobutsi EH, and Mota WF. 2015 Caracterização física, química e nutricional da pitaia-rosa de polpa vermelha. *Revista Brasileira de Fruticultura*. 37(1).

- Crane JH, andBalerdi CF. 2005. Pitaya growing in the Florida home landscape. Orlando: IFAS Extension of University of Florida.
- Donadio, LC. 2009. Pitaya. *Revista Brasileira de Fruticultura*. 31(3):637-929.
- Efraim P, Tucci ML, Pezoa-Garcia NH, Haddad R, andEberlin MN. 2006. Teores de compostos fenólicos de sementes de cacauero de diferentes genótipos. *BrazilianJournalof Food Technology*. 9(4): 229-236.
- Escribano J, Gandi'a-Herrero F, Caballero N, Pedreno, MA. 2002. Subcellular localization and isoenzyme pattern of peroxidase and polyphenol oxidase in beet root (*Beta vulgaris* L). *Journal of Agricultural and Food Chemistry*. 50:6123-6129.
- Esquivil PFC, Stintzing FC, and Carle R. 2007. Comparison of morphological and chemical fruit traits from different pitaya genotypes (*Hylocereus* sp.) grown in Costa Rica. *J. Applied Botany Food Quality*. 81:7-14.
- Franco BDGM, andLandgraf MLS. 2003. *Microbiologia dos alimentos*, Ed. Ateneu, São Paulo, Brasil. 182 pp.
- Herbach KM, Rohe M, Stintzing FC, and Carle R. 2006. Structural and chromatic stability of purple pitaya (*Hylocereuspolyrhizus* [Weber] Britton & Rose) betacyanins as affected by the juice matrix and selected additives. *Food Research International*. 39: 667-677.
- ImehU, and Khokhar S. 2002. Distribution of Conjugated and Free Phenols in Fruits: Antioxidant Activity and Cultivar Variations. *Journal of Agricultural and Food Chemistry*. 50:6301-6306.
- Jamilah B, Shu CE, Kharidah M, Dzulkifly MA, and Noranizan A. 2011. Physico-chemical characteristics of red pitaya (*Hylocereuspolyrhizus*) peel. *International Food ResearchJournal*. 18 (1): 279-286.
- Junqueira KP, Junqueira NTV, Ramos JD, andPereira AV. 2002. Informações preliminares sobre uma espécie de pitaya do Cerrado. Planaltina, DF: Embrapa Cerrados. (Documentos / Embrapa Cerrados, ISSN 1517- 5111; 62.
- Lima CA. Caracterização, propagação e melhoramento genético de pitaya comercial e nativa do Cerrado. 2013. PhD Thesis, Faculdade de Agronomia e Medicina Veterinária, Universidade de Brasília, Brasília, Brasil.
- Mahan LK, andEscott-Stump S. 2005. *Alimentos, nutrição & dietoterapia*. 11. ed. Roca: São Paulo.
- Mello FR, Bernardo C, Dias CO, Gonzaga L, Amante ER, Fett R, and Candido LMB. 2015. Antioxidant properties, quantification and stability of betalains from pitaya (*Hylocereusundatus*) peel. *Ciência Rural*. 45(2): 323-328.
- Muñoz DC, and Solano JAL. 2002. *Tablas de valor nutritivo de alimentos: Los alimentos y sus nutrientes*. McGraw Hill Interamericana: México. 203 pp.
- Natale W, Prado RM, and Mõro FV. 2005. Alterações anatômicas induzidas pelo cálcio na parede celular de frutos de goiabeira. *Pesquisa Agropecuária Brasileira, Brasília*. 40 (12):1239-1242.
- Nerd A, Tel-Zur N, and Mizrahi Y. 2002. Fruit of vineand column arcacti. In: NOBEL, PS (eds.). *Cacti: biology and uses*. UCLA, Los Angeles.
- Nunes EM, Souza ASB, Lucena CM, Silva SM, Lucena RFP, Alves CAB, and Alves RE. 2014. Pitaia (*Hylocereus* sp.): Uma revisão para o Brasil. *Gaia Scientia*. 8 (1): 90-98.
- Oliveira LA, Abreu WC, Oliveira CL, Pinto KM, Carvalho GBM, and Barcelos MFP. 2010. Composição química da pitaia vermelha (*HylocereusPolyrhizus*) e branca (*Hylocereusundatus*). XIX Congresso de Pós-graduação da UFLA, Lavras, Brasil.
- Pimentel CVMB, Francki VM, andGollucke APB. 2005. *Alimentos funcionais: introdução às principais substâncias bioativas em alimentos*. Ed. Varela: São Paulo, Brasil.
- Pinto KM, Abreu WC, Oliveira CL, Barcelos MFP, Oliveira LA, and Carvalho GBM. 2010. Caracterização física de duas variedades de pitaia. In: XIX CONGRESSO DE PÓS-GRADUAÇÃO DA UFLA, Lavras, Brasil.
- Reynerston KA, Yang H, Jiang B, Basile MB, andKennely EJ. 2008. Quantitative analysis of antiradical phenolic constituents from fourteen edible Myrtaceae fruits. *Food Chemistry*. 109 (4):883-890.
- Rufino MSM, Alves RE, Brito ES, Morais SM, Sampaio CG, andPérez-Jiménes J. 2007. Metodologia científica: determinação da atividade antioxidante total em frutas pela captura livre DPPH. *EMBRAPA Com Téc.* 127:1-4.
- Santos MRPV, Castro, JC, Mardigan LP, Watanabe R, and Clemente E. 2016. Características físico-químicas, compostos bioativos, atividade antioxidante e enzimática de frutos da pitaia (*Hylocereusundatus*). *R. Bras. Tecnol. Agroindustr.* 10 (2): 2081-2095.
- Sato STA, Ribeiro SCA, Sato MK, and Souza JNS. 2014. Caracterização física e físico-química de pitayas vermelhas (*Hylocereuscostaricensis*) produzidas em três municípios paraenses. *Journal of Bionergy and Food Science*. 1(1):46-56.
- Stintzing FC, and Carle R. 2004. Functional properties of anthocyanins and betalains in plants, food, and in human nutrition. *Trends in Food Science and Technology*. 15:19-38.
- Stintzing FC, Conrad J, Klaiberb I, Beifussb U, and Carlea R. 2004. Structural investigations on betacyanin pigments by LC NMR and 2D NMR spectroscopy. *Phytochemistry*. 65:415-422.
- Stintzing FC, Schieber A, and Carle R. 2003. Evaluation of colour properties and chemical quality parameters of cactus juices. *Eur. Food Res. Technol*. 216:303-311.
- Takata WHS. Florescimento e frutificação de pitaya vermelha com diferentes concentrações e épocas de aplicação de GA3. 2012. Masters dissertation, Universidade Estadual Paulista, Faculdade de Ciências Agrônomicas, Botucatu, Brasil.
- Vaillant F, Perez A, Davila I, Dornier M, and Reynes, M. 2005. Colorant and antioxidant properties of red pitahaya (*Hylocereus* sp.). *Fruits*. 60:1-7.
- Wall MM, and Khan SA. 2008. Postharvest quality of dragon fruit (*Hylocereus* spp.) after X-ray irradiation quarantine treatment. *HortScience*, 43:2115-2119.
- Watanabe HS, and Oliveira SL. 2014. Comercialização de frutas exóticas. *RevistaBrasileira de Fruticultura*. 6(36): 23-28.
- Waterhouse AL. 2002. Polyphenolics: determination of total phenolics. In: Wrolstad RE (eds.). *Current protocols in food analytical chemistry*. J. Wiley: New York.
- Wu LC, Hsu HW, Chen YC, Chiu CC, Lin YI, and Ho JA. 2006. Antioxidant and antiproliferative activities of red pitaya. *Food Chemistry*. 95:319-327.
- Wu SJ, and Ng LT. 2008. Antioxidant and free radical scavenging activities of wild bitter melon (*Momordica charantia* Linn. Var. abbreviate Ser.) In Taiwan. *Food Science and Technology*, 41:323-330.
- Zainoldin KH, and Baba AS. 2009. The Effect of *Hylocereuspolyrhizus* and *Hylocereusundatus* on physicochemical, proteolysis, and antioxidant activity in yogurt. *World Academy of Science, Engineering and Technology*. 60:361-366.
