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SUSTAINABLE USE AND EFFECT OF SOLVENTS ON PHYTOCHEMICAL EXTRACTION OF FIVE VULNERABLE ECONOMICALLY IMPORTANT PLANTS FROM NARAYANPUR, BASTAR, CHHATTISGARH

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ABSTRACT

Ethno-botany and ethno-medicinal investigations are now recognized as most efficient techniques for keeping informed or refocusing the data on medicinal plants reported in prior research for the possible extraction of useful bio-active compounds. Ethno-botanical research is requisite for exploring the traditional uses of plants for medicinal purposes. Several studies over the past decade have shown promising applications of phytochemicals in drug discovery and treatment. Increasingly we are observing a growing trend of applications of phytochemicals in medical science which is leading to the exponential uncontrol use of medicinal plants with economic importance. The study aims to investigate the presence of phytochemical and bioactive compounds in five medicinal plants by Soxhlet extraction method. Because of the economic and ethnomedicinal importance the selected plants are facing the status of vulnerable in Chhattisgarh state. These studies also emphasizes on bringing awareness among the folk for their sustainable use. For extraction solvents with different polarities viz., aqueous, acetone, chloroform and petroleum ether are used. Results of qualitative phytochemical screening revealed the presence of alkaloids, phenols, reducing sugar, saponins, tannins, terpenoids, flavonoids, fixed oil and fats, anthraquinones as the major bioactive compounds in all five selected plants. While cardiac glycoside was found to be absent. Maximum number of bioactive phytochemicals were extracted in the aqueous and acetone leaf extracts of *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., and *S. urens* Roxb.. While in *D. hispida* Dennst. maximum phytochemicals were observed to be present in the acetone and chloroform tuber extracts. This may be due to different solvents have been reported to have the capacity to extract different phytoconstituents depending on their solubility or polarity in the solvent. As, medicinal plants possess curative properties due to the presence of various bioactive chemical substances of different composition. Results of this study can serve as a preliminary source of knowledge for further isolation, characterization and identification of the bioactive compounds. Selected medicinal plants can also serve as a source of raw material for pharmaceutical industries as a natural source of drug designing. Dose dependent clinical trials are recommended. These plants can serve as a natural source of new drug with cheaper cost and very less side effects. Also, further research on the potential of diversity, ethnobotany, ethnopharmacology and bioactivity of plants used by local tribes is recommended to be documented for restoration of traditional knowledge of Vaid as and to bring awareness among the folk for their sustainable use and protection is being recommended.

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INTRODUCTION

Chhattisgarh renowned as the 'Herbal State', is a premier biodiversity hub with over 44% forest cover, hosting roughly 800 plus reported medicinal plant species. The state exhibits remarkable levels of endemism, particularly among plants with medicinal properties, underscoring its critical role in biodiversity conservation and the preservation of traditional healthcare practices. These plants support the livelihoods of forest dwelling communities and hold immense

economic potential for the pharmaceutical and cosmetic industries, with over 75 species highly traded for their therapeutic value. The rich biodiversity, combined with the presence of numerous tribal communities who have long relied on plants for healthcare, makes Chhattisgarh a significant centre for ethno-medicinal knowledge and practices. Tribal communities in Chhattisgarh have a deep-rooted understanding of medicinal plants utilizing them for treating various ailments. This is due to a number of reasons including arability, affordability, accessibility and low cost (Mintah *et al.* 2019). Phytochemicals are primary and secondary compounds. The term 'Phyto' derived from the Greek word which means 'Plant' are

chemical compounds naturally present in the plants attributing to positive or negative health effects. Phytochemicals are naturally occurring medicinally important chemical compounds derived or extracted from various parts of plants such as roots, stem, bark, seeds, leaves, flowers, fruits, tubers, rhizomes and even pulps. The widespread trend to humans to use plant-based medicines due to the high costs of prescription drugs and their increased side effects, there is a growing demand for many medicinal plant species in the pharmaceutical industry and traditional medicine (Chen *et al.* 2015). Medicinal plants represent promising sources for the treatment of various diseases because of their abundance in bioactive compounds with therapeutic properties. Since time immortal, or history, various ailments, and studies have revealed the presence of metabolites with anti-inflammatory, antibacterial, antiviral, antiparasitic, antidiarrheal, antioxidant, and anticancer activities. The secondary metabolites responsible for these properties include alkaloids, phenolic compounds, terpenoids, tannins, anthraquinone, cardiac glycoside, with the latter, particularly flavonoids, being the most associated with their bioactivities (Julian-Flores *et al.* 2025). The plants selected for phytochemical analysis possess different activities as mentioned in Table no. 1.

MATERIALS AND METHODS

Plant collection: Mature green and disease-free leaves of *Desmodium gangeticum* (L.) DC., *Celastrus paniculatus* Willd., *Dillenia indica* Linn., *Sterculia urens* Roxb. and tubers of *Dioscorea hispida* Dennst. were collected from Herbal Garden, Bakhrapara Kasthagar and Raoghat tropical dry deciduous forest area of Narayanpur district during their flowering season. Geographical location of Narayanpur district is 19° 43' 39" N Latitude and 81° 14' 38" E Longitude.

Preparation of extracts: The plant materials were washed thoroughly under running water, air-dried at room temperature, and crushed to fine powder using mixer-grinder. Finally, the powder was stored until it was needed for extraction.

Soxhlet extraction method: Twenty-five grams of powder was uniformly packed into a thimble prepared from Whatman No. 1 filter paper. Extraction was carried out for both aqueous and organic solvent (*viz.*, acetone, chloroform and petroleum ether) of increasing polarity. 250 ml of extracting solvent in flask is heated, and its vapours condense in condenser.

Table 1. Details of the selected five plants

Plant Details		Pharmacological activity
Botanical name	<i>Desmodium gangeticum</i> (L.) Dc.	Plant possess various biological activities such as antimicrobial, antioxidant, anti-cancer, anti-ulcer, anti-diabetic, anti-inflammatory, analgesic, anthelmintic, antipyretic, anti-amnestic, anti-secretary, anti-leishmanial, anti-implantation, cardioprotective, hepatoprotective, renal protective, psychopharmacological, immunomodulatory activities, cytoprotective, hypocholesterolemic along with it is effective as anti-writhing and central nervous system (CNS) depressant activity, improving memory and in healing different types of wounds (Mishra <i>et al.</i> 2005; Kawale <i>et al.</i> 2011-12; Rashidi and Deokule 2013; Mahajon <i>et al.</i> 2015; Tailor and Mishra 2023; Jagtap <i>et al.</i> 2025).
Common/local name	Shalparni	
Family	Fabaceae	
Habit	Shrub	
Part used	Whole plant, roots, barks	
Flowering Season	March to December	
Threat Status in India and Chhattisgarh	In India: Vulnerable (http://www.iucn.org.in) In Chhattisgarh: Vulnerable (Chandrakar 2018; http://www.iucnredlist.org ; http://www.envis.frlht.org)	
Botanical name	<i>Celastrus paniculatus</i> Willd.	Possess biological activities like antifungal, antibacterial, antiviral, antioxidant, anti-inflammatory, antidepressant, anti-arthritis, anti-fertility, analgesic, hypo-lipidemic, anti-malarial, anxiolytic, anti-aging, hypolipidemic, anti-apoptotic, gastroprotective, anti-ulcerative properties, anti-cancerous, cardiovascular, neurological disorders, learning and memory enhancer, anti-depressant, central nervous system, wound healing and immunological activity (Wang <i>et al.</i> 2005; Gnanapragasam <i>et al.</i> 2007; Bhanumathy <i>et al.</i> 2010; Arif <i>et al.</i> 2018; Dwivedi and Maurya 2018; Kumar <i>et al.</i> 2018; Kandikattu <i>et al.</i> 2021; Choudhary and Soni 2021; Baraka <i>et al.</i> 2025; Pandey <i>et al.</i> 2025).
Common/local name	Jyotishmati	
Family	Celastraceae	
Habit	Climbing shrub	
Part used	Seeds, fruits, leaves, stem, roots, barks	
Flowering Season	August to October	
Threat Status in India and Chhattisgarh	In India: Nearly Threatened (http://www.iucn.org.in) In Chhattisgarh: Vulnerable (http://www.iucnredlist.org ; Singh 2013)	
Botanical name	<i>Dillenia indica</i> Linn.	Possess activities like antimicrobial, antibacterial, antifungal, antioxidant, anti-inflammatory, anti-cancer, anti-leukemic, anti-proliferative, anti-diabetic, anti-diarrheal, anti-malarial, anti-HIV, analgesic, cytotoxic, hepatoprotective, chemoprevention and many more (Yeshwante <i>et al.</i> 2009; Endringer <i>et al.</i> 2010; Gandhi and Mehta 2013; Islam <i>et al.</i> 2013; Lima <i>et al.</i> 2014; Biswas and Pandita 2015; Kumar <i>et al.</i> 2018; Kumar <i>et al.</i> 2018; Sharma <i>et al.</i> 2022; Sahariah <i>et al.</i> 2025).
Common/local name	Chulta or Elephant apple	
Family	Dilleniaceae	
Habit	Small tree or shrub	
Part used	Fruits, leaves, aerial parts	
Flowering Season	December to April	
Threat Status in India and Chhattisgarh	In India: Narrow distribution and small population (Chaubey <i>et al.</i> 2015) In Chhattisgarh: Least concern (http://www.iucnredlist.org)	
Botanical name	<i>Sterculia urens</i> Roxb.	Possess activities like antimicrobial, antibacterial, antioxidant, anti-cancer, anti-inflammatory, anti-diabetic, antispasmodic, analgesic and diuretic properties. Also possess gastrointestinal benefits, wound healing properties, respiratory support (Gritto <i>et al.</i> 2015; Shukla and Modi 2020; Rathore <i>et al.</i> 2022; Patil <i>et al.</i> 2024).
Common/local name	Kullu	
Family	Sterculiaceae	
Habit	Tree	
Part used	Stems, barks, leaves, roots	
Flowering Season	December to January	
Threat Status in India and Chhattisgarh	In India: Critically endangered (Jadhav <i>et al.</i> 2001) In Chhattisgarh: Vulnerable (Chandrakar 2018; http://www.iucnredlist.org ; http://www.envis.frlht.org)	
Botanical name	<i>Dioscorea hispida</i> Dennst.	Possess various activities such as antimicrobial, antifungal, nematicide, antioxidant, anti-inflammatory, anti-tumor, anti-confusion, anti-phlogistic agent, anti-analgesic, anthelmintic, anti-androgenic, hypocholesterolemic, hemolytic inhibition including potential application of radioprotective phytochemicals effect, hyperglycemic, anti-hyperlipidemic activity and hemostatic properties (Murthy <i>et al.</i> 2011; Punith Kumar <i>et al.</i> 2011; Murthy <i>et al.</i> 2015; Miah <i>et al.</i> 2018; Suryowati <i>et al.</i> 2020).
Common/local name	Baichandi	
Family	Dioscoreaceae	
Habit	Shrub	
Part used	Underground tubers	
Flowering Season	August to November	
Threat Status in India and Chhattisgarh	In India: Rare (http://www.iucn.org.in) In Chhattisgarh: Vulnerable (Chandrakar 2018; http://www.iucnredlist.org ; http://www.envis.frlht.org)	

The process of extraction was continued until the solvent in thimble tube of an extractor becomes colourless. After that the solvent extracts were filtered with the help of Whatman's No.1 filter paper (Yadav and Agrawala., 2011). After filtration, the solvent was removed by evaporations in a water bath, which gave rise to a solid mass of the extract referred to as the crude extract (Muthukrishnan and Venkatachalan 2012). Further the crude extracts were stored in labelled sterile brown bottle and eppendorf tubes and kept in refrigerator at 4°C for further use. The crude extracts were subjected to phytochemical screening tests.

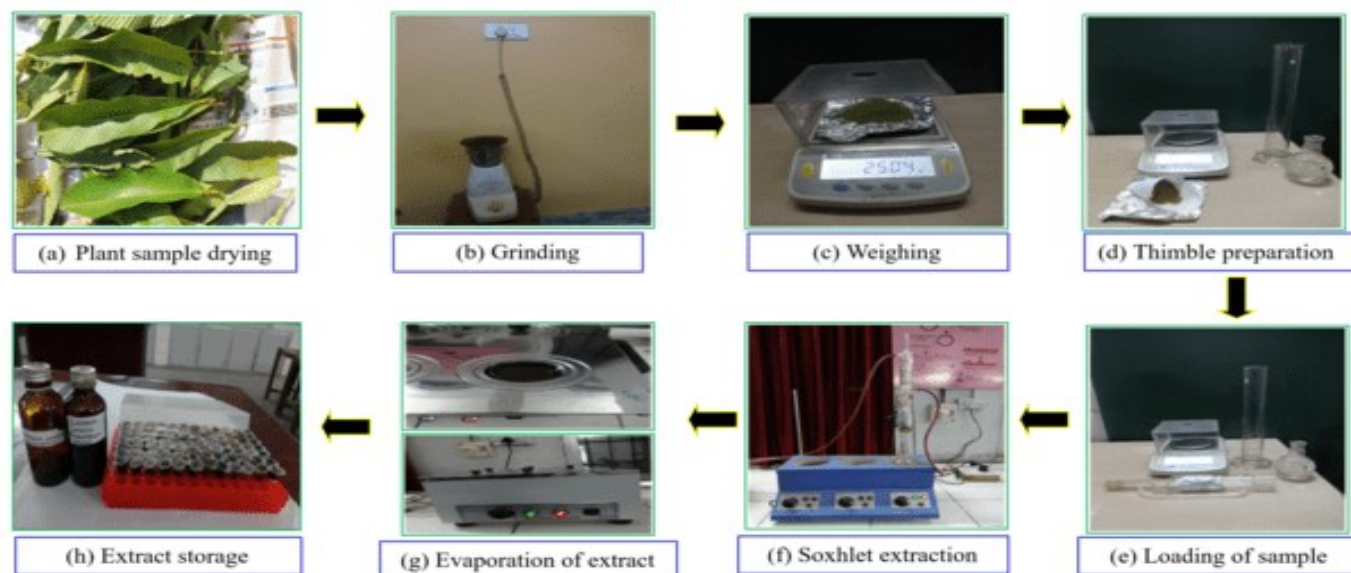


Figure 1. Preparation of plant extracts and method of Soxhlet extraction

Extraction of Secondary metabolites using different solvent extract: Qualitative phytochemical screening was based on the standard procedures described by Harborne (1973), Trease and Evans (1989), Sofowora (1993, 1996); Edeoga *et al.* (2005).

- 1) **Test for alkaloids (Mayer's test):** 6ml of extract was mixed with 6ml 1% HCl in steam bath then it was filtered. 1ml of Mayer's reagent was added. Presence of turbidity shows presence of alkaloids. Further addition of a few drops of olive oil to form an emulsion confirmed the presence of alkaloids.
- 2) **Test for phenol (Ferric chloride test):** 2ml of the crude extract was added to 4ml distilled water then few drops of 10% ferric chloride were added. Appearance of blue or green colour indicated the presence of phenol.
- 3) **Test for carbohydrate - reducing sugar (Fehling's test):** 1gm of the extract was dissolved in 10 ml distilled water. This extract was boiled with Fehling solution A and B in test tube colour changes were observed. Presence of brick red colour indicated the presence of reducing sugar.
- 4) **Test for saponins (Froth test):** 0.5gm of the extract was dissolved in 5 ml distilled water. The mixture was shaken vigorously. Formation of stable persistent froth shows the presence of saponins further addition of 6 drops of olive oil while shaking forms an emulsion confirming the presence of saponins.
- 5) **Test for tannins (Ferric chloride test):** 0.5gm of the extract was dissolved in 10 ml of distilled water a few drops of 1% ferric chloride solution was added to obtain a brownish green or blue-black precipitate confirms the presence of tannin.
- 6) **Test for terpenoids (Salkowski's test):** 0.5gm extract was dissolved in 2ml of chloroform then 3ml concentrated sulfuric acid was added. A reddish-brown colour in inter phase indicates the presence of terpenoids.
- 7) **Test for flavonoids (Ammonia reduction test):** 5ml dilute ammonia was added to 5ml extract then 5ml concentrated sulfuric acid was added formation of yellow colour shows the presence of flavonoids.

- 8) **Test for Fixed oil (Filter paper test):** A small quantity of the extracts was pressed between 2 two filter papers. Appearance of oil stain on the paper indicates the presence of fixed oils and fat.
- 9) **Test for cardiac glycosides (Keller-Killiani test):** 2.5 gm of extract was added to 2.5 ml distilled water 1 ml glacial acetic acid containing a few drops of ferric chloride was added then 0.5 ml of concentrated sulfuric acid was added presence of brown ring at the inter phase indicate the presence of deoxy sugar. A violet ring below the brown ring was observed confirming the presence of cardiac glycosides.

- 10) **Test for anthraquinone (Borntrager's test):** 2.5 gm extract was dissolved in 5ml of conc. Sulfuric acid and filtered. The filtrate was dissolved in 2.5ml of chloroform. Chloroform layer was pipetted into a tube and 0.5 ml of 10% diluted ammonia was added formation of pink red or violet colour shows the presence of anthraquinone.

RESULTS

According to preliminary phytochemical assays, the plants had different secondary metabolites. The colour change indicates the presence of different phytochemicals. And no. of plus sign indicates the presence of particular phytochemical. Results showed that more phytochemical compounds were extracted in acetone and aqueous plant extract, as compared to chloroform and petroleum ether. Except for Cardiac glycosides was found to be absent in all five plants (Table 2).

DISCUSSION

On the basis of the above findings, phytochemical screening is essential for revealing the chemical constituents that may have potential health benefits. Extraction of a phytochemical from the plant material is mainly dependent on the type of solvent used. Qualitative phytochemical screening of all five plants in the present study showed the presence of alkaloids, phenols, reducing sugar, saponins, tannins, terpenoids, flavonoids, fixed oil and fats, anthraquinones as the major bioactive compounds. These findings were similar to those of a previously carried out studies by Hill 1952; Ghosal and Bhattacharya 1972; Leete and Pinder 1972; Haq and Gomes 1974; Rahman *et al.* 1981 d; Leete and Michelson 1988; Avasthi and Tewari 1995; Mishra *et al.* 2005; Ning *et al.* 2009; Shah *et al.* 2009; Bose *et al.* 2010; Vijayalakshmi *et al.* 2011; Kawale *et al.* 2012; Vaghela *et al.* 2012; Debnath *et al.* 2012; Arora and Pandey-Rai 2012; Kalaskar *et al.* 2012; Sutte *et al.* 2012-13; Bhattacharjee 2013; Gandhi and Mehta

2013; Lima *et al.* 2014; Deodhar and Shinde 2014-15; Cui *et al.* 2014; Dat *et al.* 2015; Deodhar and Shinde 2015; Biswas and Pandita 2015; Haridas *et al.* 2016; Vepal *et al.* 2016; Lagadu and Owk 2016; Boparai *et al.* 2016; Al- Rahman 2017; El-Sherei *et al.* 2016; Dwivedi and Maurya 2018; Srivastava & Srivastava 2018; Kanoje *et al.* 2019; Sushma *et al.* 2020; Bains *et al.* 2020; Mohan *et al.* 2020; Ly *et al.* 2021; Mane *et al.* 2021; Mohan *et al.* 2021; Kanoje *et al.* 2021; Shinde *et al.* 2022; Jayasuriya *et al.* 2025; Nahar *et al.* 2025; Kabir and Lawan 2026).

Phenols present in all the extracts studied for *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., *S. urens* Roxb., *D. hispida* Dennst. Phenolic compounds are polyphenols that can be divided into flavonoids, phenolic acid, tannins, propanoid, tocopherols and phenolic diterpenes (Ali *et al.* 2008; Thagriki *et al.* 2015). Phenols are found to possess antioxidant, anti-hypertensive, blood viscosity reducer, diuretic and choleric functions. It also possesses anti-apoptosis, anti-aging, anti-carcinogenic, anti-inflammatory, anti-atherosclerosis, anti-diabetic, anti-microbial, anti-bacterial, anti-mutagenic, anti-proliferative, angiogenesis inhibitor, immune system, cardiovascular protective and also found to assist the endothelial function. Earlier researchers have also reported the use of these bioactive compounds in dye, cosmetic, food and pharmaceutical industries due to their wide therapeutic and pharmacological properties (Shui and Leong 2002; Han *et al.* 2007; Tungmunthum *et al.* 2018; Airaodion *et al.* 2019; Al-Snafi 2020; Albuquerque *et al.* 2021; Madhu *et al.* 2025).

Flavonoids present in all the extracts studied for *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., *S. urens* Roxb. whereas in *D. hispida* Dennst. present only in acetone extract. Flavonoids are hydroxylated phenolic substances synthesized by plants in response to microbial infection and possess antimicrobial substances against a wide array of microorganisms *in vitro*. Activity is most likely due to their ability to form complex with extracellular and soluble proteins and to complex with bacterial cell wall (Marjorie 1996). They also are effective antioxidants and show strong activities. It also functions as free radicals, platelet aggregation, ulcers, hepatotoxins, viruses and tumours, and protection against cytostatic (Salah *et al.* 1995; Del-Rio *et al.* 1997; Okwu 2004; Compean and Ynalvez 2014). Biological functions of flavonoids apart from its anti-oxidant and free radical scavenging capacity includes coronary heart disease prevention, hepatoprotective, osteoporosis, anti-carcinogenic properties coupled with their capacity to modulate key cellular enzyme function and neurodegenerative diseases. It also possess anti-diabetic, anti-allergic, anti-ulcer, anti-parkinson, anti-depressant, anti-hypertensive, spasmolytic, anti-inflammatory, anti-mutagenic, anti-diarrheal, anti-microbial, anti-viral properties (Barakat *et al.* 1993; Rauha *et al.* 2000; Cushnie and Lamb 2005; Ververidis *et al.* 2007; Spencer 2008; Mamadaliyeva *et al.* 2011; Kumar and Pandey 2013; Chen *et al.* 2015; Panche *et al.* 2016; Dzialo *et al.* 2016; Sulsen *et al.* 2017; Andreu *et al.* 2018; Wang and Bi 2018; Rana and Gulliya 2019; Karak 2020; Rupasinghe 2020; Wen *et al.* 2020; Mutha *et al.* 2021; Agidew 2022; Sharma *et al.* 2025).

Tannins present in *D. gangeticum* (L.) DC. (aqueous and acetone extract), *C. paniculatus* Willd. (aqueous, acetone and petroleum ether extract), *S. urens* Roxb. (acetone extract). Tannin's antioxidant property is responsible for its anti-carcinogenic and anti-mutagenic potentials by protecting cellular oxidative damage, including lipid peroxidation. The generation of superoxide radicals was reported to be inhibited by tannins and related compounds. Tannins also have the anti-inflammatory and antiulcer potency on rodents showing a strong antioxidant property for possible therapeutic applications. Tannins are effective in protecting the kidneys and also showed antimicrobial, antiviral, antibacterial, antiparasitic, antioxidant, anti-inflammatory, anti-nutritional, anti-asthmatic, anti-carcinogenic, anti-allergy, anti-hypertension and cardiovascular system-protective effects. It also possesses a wide range of pharmacological and biological activities, which include anticancer, as astringent medicine for the treatment of intestinal disorders, such as dysentery and diarrhoea, hasten the healing of wounds, antitumor and inflamed mucous membrane.

Tannins have also been reported to exert other physiological effects, such as to accelerate blood clotting, reduce blood pressure, decrease the serum lipid level, and modulate immune-responses. Analgesic and anti-inflammatory properties can be attributed to terpenoids and tannins. Apart from these tannins contribute properties of astringency i.e., faster healing of wounds and inflamed mucous membrane. Tannin has been widely used topically to sprains, bruises and superficial wounds (Chung *et al.* 1998; Okwu and Josiah 2006; Usman and Osuji 2007; Serrano *et al.* 2009; Ashok and Upadhyaya 2012; Avinish and Waman 2014; Zarin *et al.* 2016; Smeriglio *et al.* 2017; Suvanto *et al.* 2017; Britto *et al.* 2017; Ejiofor *et al.* 2018).

Terpenoids present in *D. gangeticum* (L.) DC. (aqueous extract), *C. paniculatus* Willd. (aqueous and petroleum ether extract), *D. indica* Linn. (aqueous extract), *S. urens* Roxb. (aqueous and acetone extract), *D. hispida* Dennst. (acetone extract). The presence of terpenoids in various plants was reported by various researchers (Scortichini and Rossi 1991; Tsuchiya *et al.* 1996) and that plants were widely used in herbal medicines due to the presence of terpenoids (Hayashi *et al.* 1993; Edeoga *et al.* 2005). Terpenoids have been found to be useful in the prevention and therapy of several diseases due to its anti-malarial, promote transdermal absorption, prevent and treat cardiovascular diseases and have hypoglycemic activities. Terpenoids have many potential applications, such as insect resistance, immunoregulation, antioxidant, anti-aging, neuroprotection, anti-allergenic, antispasmodic, anti-hyperglycaemic, anti-microbial, anti-bacterial, anti-viral, anti-parasitic, anti-inflammatory, anti-diabetic, anti-tumour, anti-HIV, cardio hepatoprotective effects including cancer and immune-modulatory properties. It is also found to be associated with inhibition of cholesterol synthesis and insecticidal activities and for the treatment of metabolic diseases (Mahato and Sen 1997; Fernandez *et al.* 2001; Wagner and Elmadfa 2003; Rabi and Bishayee 2009; Saleem 2009; Hill and Connolly 2012; Sulsen *et al.* 2017; Yang *et al.* 2020).

Alkaloids present in all the extracts studied for *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn. (except chloroform extract), *S. urens* Roxb., *D. hispida* Dennst. In almost all plants alkaloids are one of the most abundant and diverse groups of secondary metabolites found at 10-15% concentration. Many alkaloids are toxic and often have a pharmacological effect, which renders them as medications and recreational drugs (Manske 1965). The high abundance of alkaloids in most of the extracts confirmed the therapeutic use of the plants have been associated with medicinal uses for centuries (Nobori *et al.* 1994). Available literature tells us about the broad range of pharmacological potentials as an anti-malarial, anti-diarrheal, anti-asthmatic, anti-carcinogenic, anti-bacterial, anti-analgesic, anti-inflammatory and antispasmodic. They are found to reduce stress and depression-like symptoms (Antherden 1969; Harborne 1973; Odebiyi and Sofowora 1978; Stray 1998; Okwu and Okwu 2004).

Saponins present in all the extracts studied for *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., *S. urens* Roxb., *D. hispida* Dennst. (except petroleum ether extract). In general, saponins have been related to immunostimulatory, hypocholesterolemic, hepatoprotective, anti-tumour, anti-ulcer, antimicrobial, antibacterial, antiviral, antifungal, antiparasitic, adjuvant and cholesterol reducing activity is most pronounced. Saponins act as an expectorant and are used in the management of upper respiratory tract inflammations, atherosclerosis, diabetes, acute lung injury, cancer, and cardiovascular diseases. Medically they are used as expectorant, emetic and for the treatment of excessive salivation, epilepsy, chlorosis and migraines. Saponins are used in ayurvedic medicine as a treatment for eczema, psoriasis and for removing freckles and are also believed to be useful in the human diet for controlling cholesterol. The heart muscle causing the heart to pump more efficiently can be strengthened by treating with digitalis-type saponins. Saponins also inhibit cancer tumour growth in animals, particularly lung and blood cancers without killing normal cells. Saponins are characterized by their bitter taste and their ability to haemolyze red blood cells other characteristics including haemolytic activity, cholesterol binding and

Table 2. Results of Qualitative Analysis of Phytochemicals from Five Vulnerable Medicinal Plants

Plant species	Solvent	Phytochemical Bioactive Compounds									
		Alkaloids	Phenols	Reducing sugar	Saponins	Tannins	Terpenoids	Flavonoids	Fixed oils and fats	Cardiac glycoside	Anthraquinones
<i>Desmodium gangeticum</i> L. (DC.) (Leaves)	AQ	++	+	++	+	+	+++	+	+++	-	++
	AC	++	++	++	+	++	-	+	+++	-	-
	CF	+	+	++	+	-	-	+	+++	-	-
	PE	+	+	++	+	-	-	+	+++	-	-
<i>Celastrus paniculatus</i> Willd. (Leaves)	AQ	++	+	+	+	+	+++	+	++	-	++
	AC	+++	+	+	+	++	-	+	+++	-	-
	CF	+	+	++	+	-	-	+	++	-	-
	PE	+++	+++	+	+	+	+	+	+++	-	-
<i>Dillenia indica</i> Linn. (Leaves)	AQ	++	+	++	++	-	+++	+	++	-	++
	AC	++	+	++	+	-	-	+	++	-	-
	CF	-	+	+++	++	-	-	+	++	-	-
	PE	+	++	-	+	-	+	++	++	-	-
<i>Sterculia urens</i> Roxb. (Leaves)	AQ	+	+	++	+	-	+	+	+	-	++
	AC	+	+	++	+	++	++	+	++	-	-
	CF	+	+	++	+	-	-	+	++	-	-
	PE	+	+	-	+	-	-	++	+	-	-
<i>Dioscorea hispida</i> Dennst. (Tubers)	AQ	++	+	-	++	-	-	-	-	-	-
	AC	+	++	-	+++	-	+	++	-	-	++
	CF	+	+	+	+	-	-	-	-	-	-
	PE	+	+	-	-	-	-	-	-	-	-
Phytochemical Tests		Mayer's Test	Ferric Chloride Test	Fehling's Test	Froth Test	Ferric Chloride Test	Salkowski's Test	Ammonia Reduction Test	Filter Paper Test	Keller- Killani Test	Borntrager's Test
Inference		Presence of turbidity	Bluish green color	Brick red color	Frothing persist 15 mins	Dark blue or greenish grey Cream ppt	Bluish-green color at interphase reddish colour	Yellow color	Oil stain develops	Violet ring below brown ring interphase	Deep red or pink color of aqueous layer
Solvents: Aqueous (AQ); Acetone (AC); Chloroform (CF); Petroleum ether (PE)											
Indication: '+++' relatively a strong presence; '++' relatively moderate presence; '+' relatively low presence, '-' indicates absence.											

formation of foams in aqueous solutions (Oakenfull and Sidhu 1990; Kamel *et al.* 1991; Just *et al.* 1998; Sodipo *et al.* 2000; Okwu 2004; Moghimipour and Handali 2014; Xu *et al.* 2019). Saponins act as an immune system in plants, acting as an antibiotic to protect them against microbes and fungus. The formulation of studied plants may be helpful in the development of drugs against diseases caused by microorganisms. Saponins serve as natural antibiotics against fungal and yeast infections and help the body to fight infections and microbial invasion. Saponins have been extensively used as detergents, as pesticides and molluscicides, in addition to their industrial applications as foaming and surface-active agents and also have beneficial health effect (Shideler 1980; Chatterjee and Chakravorty 1993; Shi *et al.* 2004; Santhi *et al.* 2011; Venkataramaiah and Wudayagiri 2013; Saxena *et al.* 2013).

Reducing sugar present in all the extracts studied for *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn. (except chloroform extract), *S. urens* Roxb. (except petroleum ether extract) whereas in *D. hispida* Dennst. chloroform extract. *D. hispida* Dennst. Reducing sugar possesses

pharmacological applications such as antioxidants, anti-tumor, anti-virus, immunoregulatory, hypoglycaemic activity and plant immunity (Khatri and Baruwat Chhetri 2020).

Fixed oils and fats present in *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., *S. urens* Roxb., while absent in *D. hispida* Dennst.. It possesses biological properties for preparation of ointment. Fixed oils possess activity like anti-inflammatory, antioxidant, antihistaminic, immune-modulator, antimicrobial, analgesic effects and plant immunity. Studied plants can be used for the preparation of joint pain reliving and ointment and helps in growth and development, provides essential fatty acids and fat-soluble vitamins for the system, boost the immune system and improves food palatability (Falodun *et al.* 2008; Islam 2016; Mahboubi 2018).

Anthraquinones present in only *D. gangeticum* (L.) DC. (aqueous extract), *C. paniculatus* Willd. (aqueous extract), *D. indica* Linn. (aqueous extract), *S. urens* Roxb. (aqueous extract), *D. hispida* Dennst. (acetone extract).

Anthraquinones possess pharmaceutical properties like antimicrobial, antibacterial, antiviral, antioxidant, anti-inflammatory, anti-tumor, anti-arthritis, anti-angiogenic, anti-diabetic stilbene, cytotoxic and anti-nociceptive (Jasril 2003; Choi *et al.* 2005; Tarasiuk *et al.* 2009; Mohammed *et al.* 2013; Kosalec *et al.* 2013; Kshirsagar 2014; Park *et al.* 2016; Pang *et al.* 2020; Shami *et al.* 2020; Nam *et al.* 2021). Anthraquinones naturally occur in some plants, fungi, lichen and insects, wherein they serve as a basic skeleton for their pigments. Anthraquinones are used in the production of dyes and are also used as a laxative (Chatterjee and Chakravorty 1993; Samp 2008).

Cardiac glycosides gave negative tests for *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., *S. urens* Roxb., and *D. hispida* Dennst.. Cardiac glycosides are plant secondary metabolites that act on the contractile action of the cardiac muscle, treatment of congestive heart failure, cardiac arrhythmia and anti-cancer so these plants may become the cheapest source to develop cardio protective medicine by further pharmacological studies in future (Iyer *et al.* 2010). Glycosides are known to lower the blood pressure and also as an arrow head poisons in hunting (Nyarka and Addy 1990; Ververidiset *et al.* 2007).

CONCLUSION

On the basis of the above findings, all five selected plants viz., *D. gangeticum* (L.) DC., *C. paniculatus* Willd., *D. indica* Linn., *S. urens* Roxb., (leaves) and *D. hispida* Dennst. (tubers) are the source of secondary metabolites i.e., alkaloids, phenols, reducing sugar, saponins, tannins, terpenoids, flavonoids, fixed oil and fats, and anthraquinones. Since, bioactive compounds occurring in plant material consist of multi-components mixtures, their separation and determination still create problems. Further, to perform more precise investigations by isolation, purification and characterization of particular active compound is being suggested. The phytochemical analysis of medicinal plants is also important and have commercial interest in both research institutes and pharmaceutical companies for the manufacturing of new drugs and biofertilizers for the treatment of various plant diseases. Also, it is being suggested to take an urgent action to protect these vulnerable medicinal plants species in Chhattisgarh state and to protect and conserve the loss of natural biodiversity. Government and local tribes should take proper action to maintain its number by limiting the uncontrol use of medicinal plants. Suggestion is to follow proper agricultural practices like propagation, seed conservation and less exploitation of plants to prevent the loss and promote sustainable use of the natural resources. As herbal medicines are becoming popular and this has enhanced the ethnomedicinal investigation on herbal products. Consequently, medicinal plants are now under great pressure due to its increase demand.

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REFERENCES

1. Abd Al-Rahman AYR. 2017. A pharmacognostical study of certain Sterculia species (Family: Sterculiaceae) (thesis). Egypt. Pharmacognosy Department Faculty of Pharmacy Cairo University. *Academic Journals*.
2. Agidew M G. 2022. Phytochemical analysis of some selected traditional medicinal plants in Ethiopia. *Bulletin of the National Research Centre*. 46(87): 1-22.

3. Airaodion AI, Olatoyinbo PO, Ogbuagu U, Ogbuagu EO, Akinmolayan JD, Adekale OA, Awosanya OO, Agunbiade AP, Oloruntoba AP, Obajimi OO, Adeniji AR and Airaodion EO. 2019. Comparative assessment of phytochemical content and antioxidant potential of *Azadirachta indica* and *Parquetina nigrescens* leaves. *Asian Plant Research Journal*. 2(3): 1-14.
4. Albuquerque BR, Heleno SA, Oliveira M BPP, Barros L and Ferreira I CFR. 2021. Phenolic compounds: current industrial applications, limitations and future challenges. *Food and Function*. 12(1): 14-29.
5. Ali SS, Kasoju N, Luthra A, Singh A, Sharanabasava H, Sahuand A and Bora U. 2008. Indian medicinal herbs as sources of antioxidants. *Food Research International*. 41(1): 1-15.
6. Al-Snafi AE. 2020. Phenolics and flavonoids contents of medicinal plants, as natural ingredients for many therapeutic purposes- A review. *IOSR Journal of Pharmacy*. 10(7): 42-81.
7. Andreu L, Nuncio-Jauregui N, Carbonell-Barrachina AA, Legua P and Hernandez F. 2018. Antioxidant properties and chemical characterization of Spanish *Opuntia ficus-indica* Mill. cladodes and fruits. *Journal of the Science of Food and Agriculture*. 98(2018): 1566-1573.
8. Antherden LM. 1969. Textbook of Pharmaceutical Chemistry, 8 th edn., Oxford University Press, London, pp. 813-814.
9. Arif PT, Shaligram SS and Babasaheb KS. 2018. Pharmacological profile of *Jyotishmati* (*Celastrus paniculatus* Willd): A review. *International Journal of AYUSH*. 7(3): 901-923.
10. Arora N and Pandey-Rai S. 2012. *Celastrus paniculatus*, An endangered Indian medicinal plant with miraculous cognitive and other therapeutic properties: An overview. *International Journal of Pharma and Bio Sciences*. 3(3): 290-303.
11. Ashok PA and Upadhyaya K. 2012. Tannins are Astringent. *Journal of Pharmacognosy and Phytochemistry*. 1(3): 45-50.
12. Avasthi BK and Tewari JD. 1955. Chemical investigation of *Desmodium gangeticum*, II. Chemical constitution of the lactone. *Journal of the American Pharmaceutical Association* (Scientific ed.). 44(10): 628-629.
13. Avasthi BK and Tewari JD. 1995. A preliminary phytochemical investigation of *Desmodium gangeticum* DG. (Leguminosae). *Journal of the American Pharmaceutical Association* (Scientific ed). (Baltim). 44(10): 625-627.
14. Avinash DK and Waman SN. 2014. Phytochemical constituents of leaves of *Celastrus paniculatus* Wild: Endangered Medicinal Plant. *International Journal of Pharmacognosy and Phytochemical Research*. 6(4): 792-794.
15. Bains S, Kaur R and Sethi M. 2020. Phytochemical analysis of medicinal plants and their antimicrobial activity. *Agricultural Research*. 57(3): 444-448.
16. Baraka B B H, Rao B V and Krishnamurthy T. 2025. Synergistic effect of *Celastrus paniculatus* and *Tribulus terrestris* against chronic immobilization stress-induced memory impairment, BDNF level and neuroinflammation. *Journal of Applied Pharmaceutical Science*. 15(02): 215-223.
17. Barakat MZ, Shahab SK, Darwin N and Zahemy EI. 1993. Determination of ascorbic acid from plants. *Annal of Biochemistry*. 53: 225-245.
18. Benavente-Garcia O, Castillo J, Marin FR, Ortuno A and Del Rio JA. 1997. Uses and properties of citrus flavonoids. *Journal of Agricultural and Food Chemistry*. 45(12): 4505-4515.
19. Bhanumathy M, Chandrasekar SB, Chandur U, Somasundaram T. 2010. Phytopharmacology of *Celastrus paniculatus*: An Overview. *International Journal of Pharmaceutical Sciences and Drug Research*. 2(3): 176-181.
20. Bhattacharjee A, Shashidhara SC and Saha S. 2013. Phytochemical and ethno-pharmacological profile of *Desmodium gangeticum* (L.) DC.: A review. *International Journal of Biomedical Research*. 4(10): 507-515.
21. Biswas S and Pandita N. 2015. Phytochemical analysis and chromatographic evaluation of alcoholic extract of *Dillenia indica* Linn. leaves. *International Journal of Pharmaceutical Sciences and Research*. 6(7): 2799-2812. 13

22. Boparai A, Niazi J, Bajwa N and Singh PA. 2016. A Review update on *Dillenia indica* F. Elongata (Miq.) Miq. *Journal of Drug Delivery & Therapeutics*. 6(2): 62-70.
23. Bose U, Gunasekaran K, Bala V and Rahman AA. 2010. Evaluation of phytochemical and pharmacological properties of *Dillenia indica* Linn. leaves. *Journal of Pharmacology and Toxicology*. 5(5): 222-228.
24. Britto SJ, Mani B, Thomas S and Prabhu S. 2017. A new subspecies of *Celastrus* (Celastraceae) from the Palni hills of South India. *Taiwania*. 62(3): 311-314.
25. Chandrakar AK. 2018. Threatened medicinal plants species from Chhattisgarh. *Life Sciences International Research Journal*. 5(1): 24-28.
26. Chatterjee TK and Chakravorty A. 1993. Wound healing properties of the new antibiotics (MT81) in mice. *Indian Drugs Bombay*. 1993; 30: 450-452.
27. Chen MY, Li X, Wang L, Zhang QQ, Fang F, Miao L, Lu F and Wang H. 2015. Rapid detection of the chemicals added in traditional Chinese medicine for heat-clearing drugs by TLC-SERS. *Journal of Light Scattering*. 27: 326-331.
28. Choi SZ, Lee SO, Jang KU, Chung SH, Park SH, Kang HC, Yang EY, Cho HJ and Lee KR. 2005. Antidiabetic stilbene and anthraquinone derivatives from *Rheum undulatum*. *Archives of Pharmacol Research*. 28(9): 1027-1030.
29. Choudhary A and Soni P. 2021. Pharmacological activities of *Celastrus paniculatus* Willd. A review. *International Journal of Pharmaceutical Sciences*. 69(1): 139-144.
30. Chung KT, Wong TY, Wei CI, Huang YW and Lin Y. 1998. Tannins and Human Health: A Review. *Critical Reviews in Food Science and Nutrition*. 38(6): 421-464.
31. Compean KL and Ynalvez RA. 2014. Antimicrobial activity of plant secondary metabolites: A review. *Research Journal of Medicinal Plants*. 8(5): 204-213.
32. Cui KMRK, Merle NT and Delorina R. 2014. Studies on Phytochemical Screening and antimicrobial activity of *Dioscorea hispida* Dennst. (Kurot) tuber extract. *Journal of Applicable Chemistry*. 3(6): 2598-2601.
33. Cushnie TTP and Lamb AJ. 2005. Antimicrobial activity of flavonoids. *International Journal of Antimicrobial Agents*. 26(5): 343-356.
34. Dat NT, Luyen NT and Dang NH. 2015. Phenolic Glucosides from the leaves of *Desmodium gangeticum* (L.) DC.. *Vietnam Journal of Chemistry*. 53(2e): 69-72.
35. Debnath M, Biswas M and Nishteswar K. 2012. Evaluation of analgesic activity of different leaf extracts of *Celastrus paniculatus* (Willd.). *Journal of Advanced Pharmacy Education & Research*. 2(2): 68-73.
36. Deodhar KA and Shinde NW. 2014-2015. Phytochemical constituents of leaves of *Celastrus paniculatus* Willd: Endangered Medicinal Plant. *International Journal of Pharmacognosy and Phytochemical Research*. 6(4): 792-794.
37. Deodhar KA and Shinde NW. 2015. *Celastrus paniculatus*: Medicinal and pharmacological properties: A Review. *International Journal of Development Research*. 5(9): 5526-5531.
38. Deodhar KA and Shinde NW. 2015. *Celastrus paniculatus*: Traditional uses and Ethnobotanical study. *Indian Journal of Advances in Plant Research*. 2(1): 18-21.
39. Dwivedi V and Maurya H. 2018. A comprehensive overview of *Celastrus paniculatus* seed oil intended for the management of human ailments. *Indian Journal of Pharmaceutical and Biological Research*. 6(02): 37-42.
40. Dzialo M, Mierziak J, Korzun U, Preisner M, Szopa J and Kulma A. 2016. The potential of plant phenolics in prevention and therapy of skin disorders. *International Journal of Molecular Sciences*. 17(2): 160.
41. Edeoga HO, Okwu DE and Mbaebie BO. 2005. Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*. 4(7): 685-688.
42. Ejiofor NC, Ezeagu IE, Ayoola MB and Umera EA. 2018. Determination of the chemical composition of Avocado (*Persea americana*) seed. *Advances in Food Technology and Nutritional Science. Open Journal SE*. (2): S51-S55.
43. El-Sherei MM, Ragheb AY, Kassem M El S, Marzouk MM, Mosharafa SA and Saleh NAM. 2016. Phytochemistry, Biological activities and economical uses of the Genus *Steculia* and the related Genera: A Review. *Asian Pacific Journal of Tropical Disease*. 6(6): 492-501.
44. Endringer DC, Valadares YM, Campana PRV, Campos JJ, Guimaraes KG and Pezzuto JM and Braga FC. 2010. Evaluation of Brazilian plants on cancer chemoprevention targets in vitro. *Phytotherapy Research*. 24(6): 928-933.
45. Falodun A, Owolabi OJ and Osahon O. 2008. Physiochemical, antimicrobial and anti-inflammatory evaluation of fixed oil from *Boa constrictor*. *Acta Poloniae Pharmaceutica-Drug Research*. 65(4): 477-480.
46. Fernandez MA, de Las Heras B, Garcia-Gimenez MD and Saenz MT and Villar A. 2001. New insights into the mechanism of the anti-inflammatory triterpene lupeol. *Journal of Pharmacy and Pharmacology*. 53(11): 1533-1539.
47. Gandhi D and Mehta P. 2013. *Dillenia indica* Linn. and *Dillenia pentagyna* Roxb.: Pharmacognostic, phytochemical and therapeutic aspects. *Journal of Applied Pharmaceutical Science*. 3(11): 134-142.
48. Ghosal S and Bhattacharya SK. 1972. *Desmodium* alkaloids. Part II. Chemical and pharmacological evaluation of *Desmodium gangeticum*. *Planta Medica*. 22(4): 434-440.
49. Gnanapragasam A, Yogeeta S, Subhashini R, Ebenezer KK, Sathish V and Devaki T. 2007. Adriamycin induced myocardial failure in rats: Protective role of *Centella asiatica*. *Molecular and Cellular Biochemistry*. 294(1-2): 55-63.
50. Gritto MJ, Nandagopalan V and Doss A. 2015. Ethno-botanical study on the traditional healers in Pachamalai hills of Eastern Ghats, Tamilnadu, South India. *Journal of Medicinal Plants Studied*. 3(2): 80-85.
51. Han X, Shen T and Lou H. 2007. Dietary Polyphenols and their Biological Significance. *International Journal of Molecular Sciences*. 8(9): 950-988.
52. Haq QN and Gomes J. 1974. Arabino-galactan from the fruits of *Dillenia indica*. *Bangladesh Journal of Scientific and Industrial Research*. 19: 184-190.
53. Harborne JB. 1973. *Phytochemical Methods: A guide to modern techniques of plant analysis*. 1st Edition. Chapman and Hall Ltd 11 New Fetter Lane, London EC4P 4EE. pp. 279.
54. Haridas H, George CS, Krishnan D, Jose A and Jayachandran TP. 2016. A Review of pharmacognostic and phytochemical evaluation of plant *Desmodium gangeticum* (L.) DC.. *World Journal of Pharmaceutical Research*. 5(4): 463-471.
55. Hayashi F, Coles SK, Bach KB, Mitchell GS and McCrimmon DR. 1993. Time-dependent phrenic nerve responses to carotid afferent activation: intact vs. decerebellate rats. *American Journal of Physiology*. 265: R811-819.
56. Hill AF. 1952. *Economic Botany: A textbook of useful plants and plant products*. 2nd Edition. McGraw-Hill Book Company Inc, New York. pp.560.
57. Hill RA and Connolly JD. 2012. Triterpenoids. *Natural Product Reports*. 28(2011): 1087-1117. Islam Md. T, Streck L, Paz MFCJ, de Cadro e Sousa JM, de Alencar MVOB, de Mata AMOF, de Carvalho RM, de Oliveira Santos JV, da Silva-Junior AA, Ferreira PMP and de Carvalho Melo-Cavalcante AA. 2016. Preparation of phytol-loaded nanoemulsion and screening for antioxidant capacity. *International Medical Society*. 9(70): 1-15.
58. Islam MM, Pia RS, Sifath-E-Jahan K, Chowdhury J, Akter F, Parvin N and Akter S. 2013. Antidiarrheal activity of *Dillenia indica* bark extract. *International Journal of Pharmaceutical Sciences and Research*. 4(2): 682-688.
59. Iyer AKV, Zhou M, Azad N, Elbaz H, Wang L, Rogalsky DK. Rojanasakul Y, O'Doherty GA, and Langenhan JM. 2010. A direct comparison of the anticancer activities of Digitoxin MeON-Neoglycosides and O-Glycosides: Oligosaccharide chain length-dependent induction of caspase-9-mediated apoptosis. *American Chemical Society Medicinal Chemistry Letters*. 1(7): 326-330.
60. Jadhav SN, Ved DK, Ghatge U, Reddy KN and Reddy Ch. S. 2001. Proceedings of the workshop on Conservation Assessment and Management Planning for Medicinal Plants of Andhra

- Pradesh. FRLHT Bangalore. *Medicinal Plants Conservation Centre* (MPCC), EPTRI. Hyderabad.
61. Jagtap M A, Magdum A B, Lugade V, Kamble P S, Jayannawar D P, Nimbalkar M S and Patil. 2025. Chemo-profiling by UPLC-QTOF-MS, GC-MS/MS analysis and in vitro bioactivity assessment of *Desmodium gangeticum*. *South African Journal of Botany*. 277-293; 185.
 62. Jasril, Lajis NH, Mooi LY, Abdullah MA, Sukari MA and Ali AM. 2003. Antitumor promoting and antioxidant activities of anthraquinones isolated from the cell suspension culture of *Morinda elliptica*. *Asian Pacific of Molecular Biology and Biotechnology*. 11(1): 3-7.
 63. Julian-Flores A, Michel M R, Aguilar C N, Silva T G da, Torres-Leon C, Ascacio-Valdes JA, Sepulveda L, Aguilar-Zarate P and Chavez-Gonzalez M L. 2025. Encapsulation and digestive evaluation of infusion extracts from semi-desert Mexican plants: Phytochemical profiling and bioactivities. *Plants*. 14(22): 3448.
 64. Just MJ, Giner MC, Cuellar RMJ, Manez S, Bilia SAR and Rios JL. 1998. Anti-inflammatory activity of unusual Lupane saponins from *Bupleurum frutescens*. *Planta Medicine*. 64(05): 404-407.
 65. Kabir D and Lawan I. 2026. Qualitative phytochemical screening methods of bioactive compounds from medicinal plants. *Journal of Materials and Environmental Science*. 17(2): 251-266.
 66. Kalaskar MG, Saner SY, Pawar MV, Rokade DL and Surana SJ. 2012. Pharmacognostical investigation and physicochemical analysis of *Celastrus paniculatus* Willd. leaves. *Asian Pacific Journal of Tropical Biomedicine*. 2(3): S1232-S1236.
 67. Kamel MS, Ohtani K and Kurokawa T. 1991. Studies on *Balanites aegyptiaca* fruits, an antidiabetic Egyptian folk medicine. *Chemical and Pharmaceutical Bulletin*. 39(5):1229-1233.
 68. Kandikattu, Hemanth K, Amruta Narayanappa, Khanum Farhath, Narayana VVPC, Srinivasulu Doddaga. 2021. Phytochemical composition, pharmacological properties, and therapeutic applications of *Celastrus paniculatus*. *Current Traditional Medicine*. 7(1): 107-124.
 69. Kanoje T, Sharma K and Verma JN. 2019. Phytochemical screening of *Desmodium gangeticum* (L.) DC., *Dillenia indica* Linn. and *Dioscorea hispida*, medicinal plants of Chhattisgarh. *International Journal of Research and Analytical Reviews*. 6(2): 621-625.
 70. Kanoje T, Sharma K and Verma JN. 2021. In vitro antifungal activity and preliminary phytochemical screening of *Dillenia indica* Linn. leaf extracts from Narayanpur, Bastar, Chhattisgarh. *International Journal of Creative Research Thoughts*. 9(7): a655-a670.
 71. Karak P. 2020. Biological activities of flavonoids: An overview. *International Journal of Pharmaceutical Science and Research*. 10(4): 1567-1574.
 72. Kawale M, Saravanan R, Ankoliya S, Patel PR, Srivastava A, Gajbhiye N, Patel SL and Manivel P. 2011-12. Pharmacognostic characterization of *Desmodium gangeticum* (L.) DC.- An Ayurvedic medicinal plant. *International Journal of Pharmacognosy and Phytochemical Research*. 3(4): 113-120.
 73. Kawale M, Saravanan R, Ankoliya S, Patel PR, Srivastava A, Gajbhiye N, Patel SL and Manivel P. 2011-12. Pharmacognostic characterization of *Desmodium gangeticum* (L.) DC.- An Ayurvedic medicinal plant. *International Journal of Pharmacognosy and Phytochemical Research*. 3(4): 113-120.
 74. Khatir D and Baruwat Chhetri SB. 2020. Reducing sugar, total phenolic content, and antioxidant potential of Nepalese plants. *BioMed Research International*. 2020: 1-7.
 75. Kosalec I, Kremer D, Locatelli M, Epifano F, Genovese S, Carlucci G, Randic M and Zovko Koncic M. 2013. Anthraquinone profile, antioxidant and antimicrobial activity of bark extracts of *Rhamnus alaternus*, *R. fallax*, *R. intermedia* and *R. pumila*. *Food Chemistry*. 136(2): 335-341.
 76. Kshirsagar AD, Panchal PV, Harle UN, Nanda RK and Shaikh HM. 2014. Anti-inflammatory and antiarthritic activity of anthraquinone derivatives in rodents. *International Journal of Inflammation*. 2014: 1-12.
 77. Kumar P, Kumar S and Kumar V. 2018. Recent update on pharmacology and phytochemistry of *Dillenia indica*. *International Journal of Pharmacy and Biological Sciences*. 8(3): 805-818.
 78. Kumar S and Pandey AK. 2013. Chemistry and biological activities of flavonoids: An overview. *The Scientific World Journal*. 2013: 1-16.
 79. Kumar V, Prasher JB and Raghuwanshi S. 2018. *Dillenia indica*: An ethno-medicinal plant with high values in pharmaceutical industry. *International Journal of Advance Research in Science and Engineering*. 07(03): 441-450.
 80. Lagudu MN and Owk AN. 2016. Antimicrobial activity and phytochemicals constituents of *Desmodium gangeticum* leaves. *International Research Journal of Agricultural and Food Sciences*. 1(3): 44-52.
 81. Leete E and Michelson RH. 1988. Biosynthesis of dioscorine from trigonelline in *Dioscorea hispida*. *Phytochemistry*. 27(12): 3793-3798.
 82. Leete E and Pinder AR. 1972. Biosynthesis of dioscorine. *Phytochemistry*. 11(11): 3219-3224.
 83. Lima CC, Lemos RPL and Conserva LM. 2014. *Dilleniaceae* family: An overview of its ethnomedicinal uses, biological and phytochemical profile. *Journal of Pharmacognosy and Phytochemistry*. 3(2): 181-204.
 84. Ly HT, Truong TM, Nguyen TTH, Nguyen HD, Zhao Y and Le VM. 2021. Phytochemical screening and anticancer activity of the aerial parts extract of *Xanthium strumarium* L. on HepG2 cancer cell line. *Clinical Phytoscience*. 7(2021): 1-14.
 85. Mahajon B, Deepak M, Rema Shree AB and Remadevi R. 2015. Comparative phytochemical analysis of different parts of *Sthiraa-Desmodium gangeticum* (L.) DC. *World Journal of Pharmaceutical Research*. 4(08): 1821-1828.
 86. Mahato SB and Sen S. 1997. Advances in triterpenoid research, 1990-1994. *Phytochemistry*. 44(7): 1185-1236.
 87. Mahboubi M. 2018. Natural therapeutic approach of *Nigella sativa* (Black seed) fixed oil in management of Sinusitis. *Integrative Medicine Research*. 7(1): 27-32.
 88. Mamadaliyeva NZ, Herrmann F, El-Readi MZ, Tahrani A, Hamoud R, Egamberdieva DR, Azimova SS and Wink M. 2011. Flavonoids in *Scutellaria immaculata* and *S. ramosissima* (Lamiaceae) and their biological activity. *Journal of Pharmacy and Pharmacology*. 63(10): 1346-1357.
 89. Mane NB, Shaikh NA and Ingawale SV. 2021. Preliminary phytochemical screening of some compounds from leaf and stem of *Putranjiva roxburghii* Wall. *Journal of Pharmacognosy and Phytochemistry*. 10(1): 444-447.
 90. Manske RHF. 1965. The Alkaloids: Chemistry and Physiology. Volume 8. Indole alkaloids. Academic Press Inc., New York-London. Von R. H. F. Manske Bd. pp 159.
 91. Marjorie C. 1996. Plant products as antimicrobial agents. *Clinical Microbiology Reviews*. 12: 564-582.
 92. Miah MM, Das P, Ibrahim Y, Shajib Md. S and Rashid MA. 2018. In vitro antioxidant, antimicrobial, membrane stabilization and thrombolytic activities of *Dioscorea hispida* Dennst. *European Journal of Integrative Medicine*. 19(2018): 121-127.
 93. Mintah ST, Asafo-Agyei T, Archer MA, Junior P AA, Boamah D, Kumadoh D, Appiah A, Ocloo A, Boakye Y D and Agyare C. 2019. Medicinal Plants for Treatment of Prevalent Diseases. *Pharmacogenesis of Medicinal Plants*. Pp. 1-19.
 94. Mishra PK, Singh N, Ahmad G, Dube A and Maurya R. 2005. Glycolipids and other constituents from *Desmodium gangeticum* with antileishmanial and immunomodulatory activities. *Bioorganic & Medicinal Chemistry*. Letters. 15(20): 4543-4546.
 95. Mishra PK, Singh N, Ahmad G, Dube A and Maurya R. 2005. Glycolipids and other constituents from *Desmodium gangeticum* with antileishmanial and immunomodulatory activities. *Bioorganic & Medicinal Chemistry*. Letters. 15(20): 4543-4546.
 96. Moghimipour E, Sadaghi-Nejad B, Handali S, Ameri A, Ramezani Z and Azemi ME. 2014. In vitro screening of anti-candida activity of saponins extracted from *Glycyrrhiza glabra* and *Quillaja saponaria*. *Asian Journal of Pharmaceutical Clinical Research*. 7(1): 160-162.

97. Mohammed MMD, El-Souda SS, El-Hallouty SM and Kobayashi N. 2013. Antiviral and cytotoxic activities of anthraquinones isolated from *Cassia roxburghii* Linn. leaves. *Herba Polonica*. 59(4): 33-44.
98. Mohan P K, Krishna T P. 2021. Pharmacological profiling of *Desmodium gangeticum* (L.) DC. with special reference to soil chemistry. *Futur Journal of Pharma Science*. 7(1): 210.
99. Mohan PK, Moola AK, Kumar TS, Kumari BD R. 2020. A comprehensive review of the phytochemical and pharmacological properties of *Desmodium gangeticum* (L.) DC. *Journal of Advanced Scientific Research*. 11: 90-97.
100. Murthy GP, Kumar PTG, Suresh A, Ravishankar HG, Chandrasekhar KB and Lokesh S. 2011. Evaluation of ethanolic leaf extract of *Dioscorea hispida* Dennst. for anti-inflammatory and analgesic activities. *International Journal Pharmacy & Industrial Research*. 1(2): 83-87.
101. Murthy P, Chandrasekhar KB and Lokesh S. 2015. Phytochemical and pharmacological evaluation of ethno-medicinal plant drugs (EMP) and Tribal medicine formulation (TMF) used by Tribal practitioners for wound therapeutics in the region of Biligiriranga hills, Karnataka. *International Journal of Research in Applied, Natural and Social Sciences*. 3(7): 41-66.
102. Mutha RE, Tatiya AU and Surana SJ. 2021. Flavonoids as natural phenolic compounds and their role in therapeutics: an overview. *Future Journal of Pharmaceutical Sciences*. 7(25): 1-13.
103. Nahar L, Habibi E, Khuniad C, Kalieva K, Wang D, Arabnozari H, Chaiwut P, Sangthong S, Theansungnoen T, Nath R, Talukdar A D and Sarker S D. 2025. Bioactive phytochemicals, pharmacological, and therapeutic potential of *Dillenia indica*: A comprehensive review of current research. *Chinese Herbal Medicines*. 17(4): 628-642.
104. Nam J, Seol DW, Lee CG, Wee G, Yang S and Pan CH. 2021. Obtusifolin, an anthraquinone extracted from *Senna obtusifolia* (L.) H.S.Irwin and Barneby, Reduces inflammation in a mouse Osteoarthritis model. *Pharmaceuticals*. 14(3): 249.
105. Ning G, Tianhua L, Xin Y and He P. 2009. Constituents in *Desmodium blandum* and their antitumor activity. *Chinese Traditional and Herbal Drugs*. 40(6): 852-856.
106. Nobori T, Miura K, Wu DJ, Lois A, Takabayashi K and Carson DA. 1994. Deletion of cyclin-dependent kinase-4 inhibitor gene in multiple human cancers. *Nature*. 368(1994): 753-756.
107. Nyarka A and Addy M. 1990. Effect of aqueous extract of *Adenia cissampeloides* on blood pressure and serum analytes of hypertensive patients. *Phytotherapy Research*. 4(1): 25-28.
108. Oakenfull D and Sidhu GS. 1990. Could saponins be a useful treatment for hypercholesterolaemia. *European Journal Clinical Nutrition*. 44(1): 79-88.
109. Odebiyi OO and Sofowora EA. 1978. Phytochemical screening of Nigerian medicinal plants II," *Lloydia*. 41(3): 234-246.
110. Okwu DE and Josiah C. 2006. Evaluation of the chemical composition of two Nigerian medicinal plants. *African Journal of Biotechnology*. 5(4): 357-361.
111. Okwu DE and Okwu ME. 2004. Chemical composition of *Spondia mombin* (Linn) plant parts. *Journal of Sustainable Agriculture and the Environment*. 6(2): 30-34.
112. Okwu DE. 2004. Phytochemicals and vitamin content of indigenous species of South-Eastern Nigeria. *Journal of Sustainable Agriculture and the Environment*. 6(1): 30-37.
113. Panche AN, Diwan AD and Chandra SR. 2016. Flavonoids: an overview. *Journal of Nutritional Science*. 5(e47): 1-15.
114. Pandey P K, Nandi M K and Babu R. 2025. Exploring the therapeutic potential of *Celastrus paniculatus*: An inclusion review. *International Journal of creative research thoughts*. 13(8): c530-c545)
115. Pang Xu, Zhao JY, Liu N, Chen MH, Zheng W, Zhang J, Chen S, Yu LI-Y and Ma BP. 2020. Anthraquinone analogues with inhibitory activities against influenza, a virus from *Polygonatum odoratum*. *Journal of Asian Natural Products Research*. 23(8): 717-723.
116. Park JG, Kim SC, Kim YH, Yang WS, Kim Y, Hong S, Kim KH and Yoo BC, Kim SH, Kim JH and Cho JY. 2016. Anti-inflammatory and antinociceptive activities of Anthraquinone-2-carboxylic acid. *Mediators of Inflammation*. 2016(2): 1-12.
117. Patil P B, Upasani MP, Patil D D, Patil R D, Patil P K and Patil G D. 2024. *Sterculia urens* Roxb.: Bridging tribal livelihoods and industrial innovation. *International Journal of Innovative Research in Technology*. 11(2): 341-345.
119. Punith Kumar TG, Panduranga Murthy G, Suresh A, Suresh V, Senthil Kumar N and Raviashankar HG. 2011. Evaluation of antitumor activity and antioxidant status in *Dioscorea hispida* Dennst. leaves on Ehrlich Ascites Carcinoma in Swiss Albino mice. *International Journal of Drug Development and Research*. 3(2): 203-210.
120. Rabi T and Bishayee A. 2009. Terpenoids and breast cancer prevention. *Breast Cancer Research and Treatment*. 115(2): 223-239.
121. Rahman ML, Haq QN and Ahmed M. 1981 d. Structural studies on polysaccharides of fruits of *Dillenia indica* Linn. Part IV. Partial hydrolysis and Smith degradation of the acidic polysaccharides. *Bangladesh Journal of Science Indian Research*. 16: 125-133.
122. Rajapakshe R M T N, Jayasuriya W J A B N; Herath H M D R, Krunarathna K A A U and Dahanayake J M. 2025. Comparative evaluation of anti-inflammatory activity and acute toxicity of *Alysicarpus vaginalis* (L.) DC. and *Desmodium gangeticum* (L.) DC. whole plants and their roots. *Traditional Medicine Research*; 10(9): 55.
123. Rana AC and Gulliya B. 2019. Chemistry and pharmacology of flavonoids- A review. *Indian Journal of Pharmaceutical Education and Research*. 53(1): 8-20.
124. Rashidi M and Deokule SS. 2013. Study on Biodeterioration of some chemical constituents in fresh and market drug *Desmodium gangeticum* DC. storage. *European Journal of Experimental Biology*. 3(1): 161-165.
125. Rathod K, Mayur R L, Pajapati D, Dhaka R, Jaliya R L, Jha S S, Desai B S and Rot J. 2022. *Sterculia urens*: Traditionally important medicinal tree. *Journal of Medicinal Plants Studies*. 10(1): 23-26.
126. Rauha JP, Remes S, Heinonen M, Hopia A, Kahkonen M, Kujala T, Pihlaja K, Vuorela H and Vuorela P. 2000. Antimicrobial effects of finnish plant extracts containing flavonoids and other phenolic compounds. *International Journal of Food Microbiology*. 56(1): 3-12.
127. Rupasinghe HPV. 2020. Special Issue "Flavonoids and their disease prevention and treatment potential": *Recent advances and future perspectives*. *Molecules*. 25(20): 1-7.
128. Saha MR, Alam Md. A, Hasan SM R, Akter R, Hossain Md. M, Mazumder EH and Rana Md. S. 2009. In vitro antioxidant activity of the leaves of *Dillenia indica*. *Oriental Pharmacy and Experimental Medicine*. 9(4): 277-284.
129. Sahariah P, Bora D, Malik S, Bhan S, Bruah TJ, Rustagi S, Thapliya S Talukdar N and Deshwal RK. (2025). Pharmacognostic evaluation of *Dillenia indica* L. fruit for its in-vitro and in-vivo antioxidant potential and investigation of bioactive phytochemicals using GC-MS and in-silico analysis. *Comparative Clinical Pathology*. 34(6)-1283-1294.
130. Salah W, Miller N, Pagaugé G, Tyburg G, Bolwell E, Rice E and Evans C. 1995. Polyphenolic flavonols as scavengers of aqueous phase radicals and as chain-breaking antioxidants. *Archives of Biochemistry and Biophysics*. 322(2): 339-346.
131. Saleem M. 2009. Lupeol, a novel anti-inflammatory and anti-cancer dietary triterpene. National Institute of Health, *Public Access. Cancer Letters*. 285(2): 109-115.
132. Samp JC. 2008. A comprehensive mechanism for anthraquinone mass transfer in alkaline pulping (Thesis). Georgia Institute of Technology. pp30.
133. Santhi R, Lakshmi G, Priyadarshini AM and Anandaraj. 2011. Phytochemical screening of *Nerium oleander* leaves and *Momordica charantia* leaves. *International Research Journal Pharmacy*. 2(1): 131-135.

134. Sarkar B, Biswas P, Adhikari S, Roy D, Behera BK, Madhu N R and Erfani H. 2025. Pharmacological uses of new bioactive compounds from medicinal plants. *Technology and Innovation for Inclusive Development*. 2: 131-171.
135. Saxena M, Saxena J, Nema R, Dharmendra S and Gupta A. 2013. Phytochemistry of medicinal plants. *Journal of Pharmacognosy and Phytochemistry*. 1(6): 168-182.
136. Scortichini M and Rossi MP. 1991. Preliminary in vitro evaluation of the antimicrobial activity of terpenes and terpenoids towards *Erwinia amylovora* (Burrill) Winslow. *Journal of Applied Bacteriology*. 71(2): 109-112.
137. Serrano J, Puupponen-Pimia R, Dauer A, Aura AM and Saura-Calixto F. 2009. Tannins: Current knowledge of food sources, intake, bio-availability and biological effects. *Molecular Nutrition & Food Research*. 2(S2): S310-S329.
138. Shami AM M, Saadedin SM K and Hasan S F. 2020. Antibacterial and antioxidant effects of anthraquinones fraction from *Clerodendron inerme* leaves. *International Journal of Research Pharmaceutical Sciences*. 11(3): 3151-3157.
139. Sharma B P, Rani J J, Vimala, Sable P D, Kumar S, Marndi S and Das K. 2025. Bioactive compounds of medicinal plants: Structure, function, and bioactivity. 9(4): 34-50.
140. Sharma G N, Kaur H, Shrivastava B and Arora S C. 2020. A review from historical to current *Celastrus paniculatus*. *International Journal of Pharmacy and Pharmaceutical Science*. 12(8): 15-20.
141. Sharma I, Kaushik S and Bhatt B. 2022. Pharmacological and phytochemical studies of *Dillenia indica*: A review. *International Journal of Biology, Pharmacy and Allied Sciences*. 10(6): 2663-2677.
142. Shi J, Arunasalam K, Yeung D, Kakuda Y, Mittal G and Guelph JY. 2004. Saponins from edible legumes: Chemistry, processing, and health benefits. *Journal of Medicinal Food*. 7(1): 67-78.
143. Shideler H. 1980. *Yucca*; The food supplement that helps prevent and treat arthritis and high blood pressure. *Arthritis News Today*. 2: 6-8. Shinde T S, Jadhav R S and Vikhe D N. 2022. Pharmacological study and phytochemical study of *Celastrus paniculatus* seed. *Research Journal of Science and Technology*. 14(2): 111-114.
145. Shui G and Leong LP. 2002. Separation and determination of organic acids and phenolic compounds in fruit juices and drinks by HPLC. *Journal of Chromatography Analysis*. 977(1): 89-96.
146. Shukla A and Modi N. 2020. India's globally important *Sterculia urens* Roxb. in crisis: Critical insights into conservation, prospects and challenges. *Indian Journal of Natural Sciences*. 10(61): 27174-27182.
147. Singh AK. 2013. Probable agricultural biodiversity heritage sites in India: XV. The Bastar region. *Asian Agri-History*. 17(1): 3-24.
148. Smeriglio A, Barreca D, Bellocchio E and Trombetta D. 2017. Proanthocyanidins and hydrolysable tannins: occurrence, dietary intake and pharmacological effects. *British Journal of Pharmacology*. 174(11): 1244-1262.
149. Sodipo OA, Akinyi JA and Ogunbamosu JU. 2000. Studies on certain characteristics of extracts of bark of *Pansynstalia macruceras* (K schemp) pierre Exbeille. *Global Journal of Pure and Applied Sciences*. 6: 83-87.
150. Sofowora A. 1993. Medicinal Plants and Traditional Medicine in Africa. Spectrum Books Ltd, Ibadan, Nigeria, pp. 191-289.
151. Sofowora A. 1996. Medicinal Plants and Traditional Medicine in Africa. Spectrum Book Ltd., Ibadan, 100-107.
152. Spencer JPE. 2008. Flavonoids: Modulators of brain function. *The British Journal of Nutrition*. 99 E Suppl 1(E-S1): ES60-ES77.
153. Srivastava P and Srivastava G. 2018. Pharmacological and phytochemical screening of *Desmodium gangeticum* and *Moringa oleifera*. *Research Journal of Chemistry and Environment*. 22(5): 6-10.
154. Stray F. 1998. The Natural Guide to Medicinal herbs and Plants. Tiger Books International, London, pp. 12-16.
155. Sulsen V, Lizarraga E, Mamadalieva NZ and Lago JHG. 2017. Potential of terpenoids and flavonoids from Asteraceae as anti-inflammatory, antitumor, and antiparasitic agents. *Evidence-Based Complementary and Alternative Medicine*. 2017: 1-2.
156. Sulsen V, Lizarraga E, Mamadalieva NZ and Lago JHG. 2017. Potential of terpenoids and flavonoids from Asteraceae as anti-inflammatory, antitumor, and antiparasitic agents. *Evidence-Based Complementary and Alternative Medicine*. 2017: 1-2.
157. Suryowati T, Sirait RH, Siagian FE and Nursyam M. 2020. Bioactive compound impacting the metabolism and antibacterial activity of *Gadung tuber* (*Dioscorea hispida* Dennst). *Journal of Physics: Conference Series*. 1665(2020): 012030.
158. Sushma BK, Ashalatha KS, Ray P and Raveesha HR. 2020. Histochemical and phytochemical analysis of medicinally important plants. *European Journal of Medicinal Plants*. 30(4): 1-13.
159. Sutttee A, Bhandari A, Bais CS and Sharma A. 2012-13. Pharmacognostical and phytochemical evaluation of *Celastrus paniculata*. *International Journal of Pharmacognosy and Phytochemical Research*. 4(4): 227-233.
160. Suvanto J, Nohynek L, Seppanen-Laakso T, Rischer H, Salminen JP and Puupponen-Pimia R. 2017. Variability in the production of tannins and other polyphenols in cell cultures of 12 Nordic plant species. *Planta*. 246(2017): 227-241.
161. Tailor V and Mishra S. 2023. *Desmodium gangeticum*: a medicinal plant of India. *Medicinal Fabaceae of India*. ISSN: 9-17.
162. Tarasiuk J, Stefanska B, Plodzich I, Tkaczyk-Gobis K, Seksek O, Martelli S, Garnier-Suillerot A and Borowski E. 2009. Anthrax pyridones, a novel group of antitumour non-cross resistant anthraquinone analogues. Synthesis and molecular basis of the cytotoxic activity towards K562/DOX cells. *British Journal of Pharmacology*. 135(6): 1513-1523.
163. Thagriki D, Dahiru D, Yaduma GW. 2015. In vitro antioxidant activity of aqueous root extract of *Parkia biglobosa*, *Anogeissus leiocarpus* and *Moringa oleifera* on erythrocytes exposed to oxidative stress. 3(3): 9-11.
164. Trease GE and Evans WC. 1989. Pharmacognosy. 13th Edition. Bailliere, London. 882.
165. Trease GE and Evans WC. 1998. Pharmacognosy. 11th Edition. Bailliere Tindall Ltd., London. 45-75.
166. Tsuchiya HM, Sato T, Miyazaki T, Fujiwara S and Tanigaki S. 1996. Comparative study on the antibacterial activity of phytochemical flavanones against methicillin-resistant *Staphylococcus aureus*. *Journal of Ethnopharmacology*. 50(1): 27-34.
167. Tungmunthum D, Thongboonyou A, Pholboon A and Yangsabai A. 2018. Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: An overview. *Medicines*. 5(93): 1-16.
168. Usman H and Osuji JC. 2007. Phytochemical and in vitro antimicrobial assay of the leaf extract of *Newbouldia laevis*. *African Journal of Traditional Complementary and Alternative Medicines*. 4(4): 476-480.
169. Vaghela BD, Patel BR and Pandya PN. 2012. A comparative pharmacognostical profile of *Desmodium gangeticum* DC. and *Desmodium laxiflorum* DC. *Ayurvedic Pharmaceutical Standardisation*. 33(4): 552-556.
170. Vedpal SP, Dhanabal, Dhamodaran P, Chaitanya MVNL, Duraiswamy B, Jayaram U and Srivastava N. 2016. Ethnopharmacological and phytochemical profile of three potent *Desmodium* Species: *Desmodium gangeticum* (L.) DC., *Desmodium triflorum* Linn. and *Desmodium triquetrum* Linn. *Journal of Chemical and Pharmaceutical Research*. 8(7): 91-97.
171. Vedpal SP, Dhanabal, Dhamodaran P, Duraiswamy B, Chaitanya MVNL, Jeyaprakash MR and Unni J. 2016. Pharmacognostical characterization, phytochemical screening and Finger print profile of the plant *Desmodium gangeticum* DC. *International Journal of Pharmacognosy and Phytochemical Research*. 8(8): 1271-1277.
172. Venkatachalam U and Muthukrishnan S. 2012. Free radical scavenging activity of ethanolic extract of *Desmodium gangeticum*. *Journal of Acute Medicine*. 2(2): 36-42.
173. Venkataramaiah CH and Wudayagiri R. 2013. Phytochemical screening of bioactive compounds present in the seed of *Celastrus*

- paniculatus: Role in traditional medicine. Seasons. *Indo American Journal of Pharmaceutical Research*. 3(11): 9104-9111.
174. Ververidis F, Trantas E, Douglas C, Vollmer G, Kretzschmar G and Panopoulos N. 2007. Biotechnology of flavonoids and other phenylpropanoid-derived natural products. Part 1: Chemical diversity, impacts on plant biology and human health. *Biotechnology Journal*. 2(10): 1214-1234.
 175. Vijayalakshmi G, Deepti K, Arjuna Rao PV and Lakshmi KVSS. 2011. Phytochemical evaluation and antimicrobial activity of crude extracts of *Desmodium gangeticum* DC.. *Journal of Pharmacy Research*. 4(7): 2335-2337.
 176. Wagner KH and Elmadfa I. 2003. Biological relevance of terpenoids: Overview focusing on mono-di and tetraterpenes. *Annals of Nutrition and Metabolism*. 47(34): 95-106.
 177. Wang TY and Bi KS. 2018. Bioactive flavonoids in medicinal plants: Structure, activity and biological fate. *Asian Journal of Pharmaceutical Sciences*. 13(1): 12-23.
 178. Wang XS, Liu L and Fang JN. 2005. Immunological activities and structure of pectin from *Centella asiatica*. *Carbohydrate Polymers*. 60(1): 95-101.
 179. Web: <http://Envis.Frlht.Org> - Envis Centre on Conservation of Medicinal Plants, Frlht, Bangalore <Http://Frlhtenvis.Nic.In> (22 September 2010)
 180. Web: <http://iucn.org.in>
 181. Web:<http://www.iucnredlist.org>
 182. Wen K, Fang X, Yang J, Yao Y, Nandakumar KS, Salem ML and Cheng K. 2020. Recent research on flavonoids and their biomedical applications. *Current Medicinal Chemistry*. 27: 1-25.
 183. Xu C, Wang W, Wang B, Zhang T, Cui X and Li N. 2019. Analytical methods and biological activities of *Panax notoginseng* saponins: Recent trends. *Journal of Ethnopharmacology*. 236: 443-465.
 184. Yadav RNS and Agarwala M. 2011. Phytochemical analysis of some medicinal plants. *Journal of Phytochemistry*. 3(12): 10-14.
 185. Yang W, Chen Xu, Li Y, Guo S, Wang Z and Yu X. 2020. Advances in pharmacological activities of terpenoids. *Natural Product Communications*. 15(3): 1-13.
 186. Yeshwante SB, Juvekar AR, Nagmoti DM, Wankhede SS, Shah AS, Pimprikar RB and Saindane DS. 2009. Anti-inflammatory activity of methanolic extracts of *Dillenia indica* L. leaves. *Pharmacology*. 1(1): 63-66.
 187. Zarin MA, Wan HY, Isha A and Armania N. 2016. Antioxidant, antimicrobial and cytotoxic potential of condensed tannins from *Leucaena leucocephala* hybrid-Rendang. *Food Science and Human Wellness*. 5(2): 65-75
